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Chemoenzymatic and enantioselective assembly of the $(1\alpha,3a\beta,6\alpha,7a\beta)$ octahydro-1,6-methano-1H-indene framework associated with 2-isocyanoallopupukeanane: validation of a new synthetic strategy and the identification of enantiomeric switching regimes

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ABSTRACT

The octahydro-1,6-methano-1H-indene framework associated with the marine sesquiterpenoid 2-isocyanoallopupukeanane (1) has been prepared in enantiomerically pure form from the cis-1,2-dihydrocatechol 8 using Diels-Alder cycloaddition, oxa-di- π -methane rearrangement and intramolecular enolate alkylation steps as the key bond-forming events. Three distinct strategies for employing such sequences in the selective synthesis of either enantiomeric form of the target framework have been identified.

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1. Introduction

In [1](#page-11-0)991 Fusetani et al. reported¹ on the elucidation, using 2D NMR spectroscopic techniques, of the structure of 2-isocyanoallopupukeanane (1), a sesquiterpenoid isonitrile isolated from two specimens of a Phyllidia pustulosa species of nudibranch collected off Hachijo-jima Island, Japan. Such isocyano-containing species, which include the framework-isomeric natural products 2-isocyanopupukeanane (2) ,² 9-isocyanopupukeanane (3)^{[2](#page-11-0)} and 9-isocyanoneopupukeanane (4),³ are found in the secretions of a variety of marine molluscs, where they are presumed to play a defensive role, and are known to be sequestered from the sponge diet of these creatures.^{2c} The origins of the isocyano-function have been the subject of various studies 4.5 including a number conducted by one of us $(M.J.G.)$ ⁵

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As part of a program directed towards developing a comprehensive understanding of the biogenesis of compound 1 we sought to establish methods for synthesising, in either enantiomeric form, $⁶$ the</sup> associated octahydro-1,6-methano-1H-indene framework incorporating relevant functionality, especially those that would allow for ready installation of isocyano and related groups at the 2-position. In contrast to the situation with compounds 2 and $3⁷$ $3⁷$ $3⁷$ there have been few studies concerned with the total synthesis of compound 1 or its enantiomer[.8,9](#page-11-0) Indeed, the single reported total synthesis of the title natural product was described by Ho et al. in $1999⁸$ $1999⁸$ $1999⁸$ and only provided the racemic modification of the target. More recently (2006), Srikrishna and Satyanarayana have communicated⁹ a biogenetically patterned and enantiospecific synthesis of 'allopupukeanones' from 6-methylcarvone that involves, as the key step, the acid-induced Wagner-Meerwein rearrangement of a pupukeanyl cation to the corresponding allopupukeanane species. However, the extension of such chemistry to the preparation of natural product 1 has not been reported thus far.

The approach to the tricyclic framework of ent-2-isocyanoallopupukeanane (ent-1) that we have pursued is outlined in [Figure 1,](#page-1-0) and this incorporates several key transformations used during the course of our development of total syntheses of various triquinane-type sesquiterpenes including hirsutene, hirsutic acid, complicatic acid and phellodonic acid.¹⁰ A pivotal aspect of the present work was the recognition that the framework of target ent-1 (and 1) is

Figure 1. Retrosynthetic analysis of ent-2-isocyanoallopupukeanane (ent-1).

a 1,5-ethano-bridged diquinane and that this could, therefore, be assembled from a precursor of the general form 5 through its subjection to reductive cleavage of the carbonyl-conjugated cyclopropane moiety and in situ intramolecular alkylation of the ensuing enolate by the C5-appended and endo-orientated side-chain bearing a leaving group at its terminus. Conventional functional group interconversions (FGIs) of the OP and carbonyl groups within the anticipated product of this sequence should then deliver ent-2-isocyanoallopupukeanane (ent-1) and related compounds. It was expected that compound 5 could, in turn, be obtained via a photochemically promoted oxa-di- π -methane rearrangement¹¹ of the disubstituted bicyclo^[2.2.2]octenone **6** followed by or, if necessary, preceded by manipulation of the associated vinyl group so as to establish the relevant functionality on the side-chain of compound 5. Access to compound 6 was expected to be gained via conventional manipulations of ketone 7, versions of which we have obtained previously through Diels-Alder cycloaddition $reactions$ between α -chloroacrylonitrile and hydroxy-protected forms of the cis-1,2-dihydrocatechol 8^{12} 8^{12} 8^{12} Starting material 8 is readily obtained in multi-gram quantities and enantiomerically pure form (>99.8% ee) through the whole-cell biotransformation of toluene using a genetically engineered micro-organism Escherichia coli JM109 (pDTG601) that over-expresses the responsible enzyme, viz. toluenedioxygenase (TDO)[.13](#page-11-0) In an overall sense, then, there are three critical chemical sequences associated with the implementation of the proposed synthetic plan, namely the Diels-Alder cycloaddition process leading to compounds of the general form 7 and the manipulation of such adducts to give bicyclo[2.2.2]octenone 6, the photochemical rearrangement of the latter to give, after appropriate FGIs, the cyclopropa-fused diquinane 5 and, finally, a reductive alkylation process leading to the complete tricyclic framework associated with target ent-1. Each of these pivotal steps is discussed separately in the following sections as is the capacity to adapt the reported chemistry to the synthesis of either enantiomeric form of 2-isocyanoallopupukeanane, viz. 1 and/or ent-1.

2. Results and discussion

2.1. Synthesis of the substrate for the oxa-di- π -methane rearrangement reaction

The first critical chemical sequence associated with the present work started [\(Scheme 1\)](#page-2-0) with the engagement of the well known^{[14](#page-11-0)} acetonide derivative, 9, of cis-1,2-dihydrocatechol 8 in a Diels-Alder cycloaddition reaction with α -chloroacrylonitrile. Hydrolysis of the ensuing mixture of epimeric α -chloronitriles^{[12](#page-11-0)} to give the previously reported ketone 10^{12} 10^{12} 10^{12} was achieved most effectively using a modification of conditions originally reported by Evans et al.¹⁵ As a prelude to introducing the vinyl group required in a photochemical substrate of the general form 6, the enolate anion derived by deprotonation of ketone 10 was treated, in diethyl ether, with 2 mol equiv of Mander's reagent^{[16](#page-11-0)} and thus affording the b-ketoester 11 in 67% yield. Reduction of the latter with sodium borohydride in ethanol then gave β -hydroxyester 12 and its stereoisomers C9-epi-12 and C8,C9-di-epi-12 in 82% combined yield.^{[17](#page-11-0)} The structure of compound 12 was confirmed by singlecrystal X-ray analysis (see [Experimental section\)](#page-4-0).

Compound C9-epi-12, the predominant product of the reduction process, could be converted into isomer 12 (67% yield at 73% conversion) upon treatment with the weakly nucleophilic base DBU. Subjection of a mixture of compounds 12 and C8,C9-di-epi-12 to mesylation under the Crossland–Servis conditions¹⁸ then gave the corresponding mixture of mesylate 13 and isomer C8,C9-di-epi-13. Treatment of the latter mixture with DBU in hot (72 $^{\circ}$ C) benzene overnight resulted in elimination of the elements of methanesulfonic acid and formation of the unsaturated ester 14 (92%). Reaction of compound 14 with sodium borohydride in methanol/THF then gave a ca. 1:1 mixture of ester 15 and its epimer C9-epi-15 (81% combined yield) that could be separated from one another by conventional flash chromatographic techniques. Interestingly, treatment of compound C9-epi-15 with sodium methoxide in methanol at 0 to 70 \degree C for 24 h afforded a ca. 3:1 mixture of the starting material and epimer 15 (89% combined yield) and thus demonstrating that additional quantities of the desired isomer (15) could be generated from the unwanted one. Treatment of ester 15 with DIBAL-H in dichloromethane/hexane at -78 °C for 1.25 h afforded a chromatographically separable mixture of alcohol 16 (57%) and aldehyde 17 (30%). The former product could be oxidised to the latter in 51% yield using the Parikh-Doering protocol.¹⁹ Additional quantities of aldehyde **17** could also be obtained by reducing ester C9-epi-15 to aldehyde C9-epi-17 and then epimerising the latter with DBU (47% yield of compound 17 over the two steps). Wittig-type methylenation of aldehyde 17 then gave the required olefin 18 in 60% yield. Cleavage of the acetonide unit within the latter compound was achieved by exposing the substrate to activated DOWEX-50 resin in refluxing aqueous methanol for 6 days. By this

means the diol 19 was obtained in 66% yield. So, despite the harsh conditions required to effect cleavage of the acetonide group, there appeared to be no complications arising from isomerisation of the terminal olefin into a more stable internal position. Selective oxidation of the hydroxyl group remote from the bridgehead methyl group within diol 19 could be achieved using the sterically demanding oxammonium salt derived from reaction of 4-AcNH-TEMPO with p-TsOH \cdot H₂O²⁰ and the ensuing acyloin **20** (81%) was immediately subjected to O-benzoylation using benzoyl chloride in the presence of 4-(N,N-dimethylamino)pyridine (DMAP) and triethylamine. By such means the keto-ester 21 was obtained in quantitative yield.

While in the original plan [\(Fig. 1](#page-1-0)) deletion of the O-benzoyl group within keto-ester 21 would now be required in order to produce the photo-substrate 6 , our previous studies^{[10](#page-11-0)} had shown that systems related to the former compound readily engage in oxa -di- π -methane rearrangement reactions. Accordingly, and given the capacity to delete the OBz group at a later stage in the synthesis, compound 21 was selected as the substrate to be used in studies of the photochemical rearrangement process. Details are presented in the following section.

2.2. The oxa-di- π -methane rearrangement reaction and chemical manipulation of the photo-product

Following procedures developed earlier,¹⁰ a solution of compound 21 in acetone containing acetophenone (triplet sensitiser) was subjected to irradiation with a medium pressure mercury-vapour lamp at 15 $\mathrm{^{\circ}C}$ for 4.5 h [\(Scheme 2\)](#page-3-0). By such means a ca. 1:1 mixture of the C4-epimeric forms of compound 22 was obtained, albeit in a disappointing 32% combined yield. This low yield could be attributed to interference from the vinyl group associated with substrate 21, especially the potential for this moiety to participate in a competing oxa-di- π -methane rearrangement process. The formation of the C4-epimeric forms of product 22 from a single epimeric form of precursor 21 presumably arises from a secondary photochemical process involving photo-enolisation and/or Nor-rish-type 1 reactions^{[10](#page-11-0)} of the primary photo-product. The driving force for the epimerisation of the primary photo-product is the migration of the O-benzoyl unit from the exo-face to the endo-face of the cyclopropa-fused diquinane framework and thereby relieving steric interactions with the adjacent methyl group.

The chemical manipulation of photo-product 22 so as to generate a substrate for examination of the proposed intramolecular enolate alkylation step [\(Fig. 1\)](#page-1-0) involved initial reductive removal of the Obenzoyl residue. This was best accomplished by treating a methanolic solution of compound 22 with 2.2 mol equiv of samarium(II) diiodide at -78 °C for 5 min.²¹ The ensuing unsaturated ketone 23 (54%) was subjected to olefin dihydroxylation using the UpJohn conditions²² and thereby affording an inseparable and 1:1 mixture of the diastereoisomeric forms of the product diol 24 in 48% combined yield. Attempts to improve upon this outcome, in terms of both yield and diastereoselectivity, by using Sharpless asymmetric

dihydroxylation protocols^{[23](#page-11-0)} either gave no reaction at all (with ADmix- β) or resulted in a 1:1:1 mixture of the starting material and the product diols (with AD-mix- α). The selective tosylation of the primary hydroxy group within compound 24 was readily accomplished using p-toluenesulfonyl chloride in the presence of dibutyltin oxide²⁴ and thereby affording the desired epimeric mono-tosylates 25 in 77% combined yield. O-Benzoylation of the secondary hydroxyl groups within the epimeric forms of compound 25 was effected under standard conditions (BzCl, DMAP, $Et₃N$) and the co-formed bis-esters 26 and 27 were readily separated from one another by conventional chromatographic techniques and thereby obtained in yields of 40% and 55%, respectively. The illustrated stereochemistries assigned to products 26 and 27 follow from a single-crystal X-ray analysis of a derivative of the former compound (vide infra).

2.3. Completion of the synthesis through intramolecular enolate alkylation

With compounds 26 and 27 in hand studies of the validity of the proposed reductive-cleavage/intramolecular enolate alkylation chemistry (see conversion $5\rightarrow 1$, [Fig. 1](#page-1-0)) could begin. The initial experiments simply involved treating each of substrates 26 and 27 with 1.2 mol equiv of samarium(II) diiodide in THF/methanol at -78 to 18 °C for periods of up to 8 h (Scheme 3).^{[25](#page-11-0)} However, after quenching the reaction mixtures and then subjecting them to work up, only the products of cyclopropane ring-cleavage, viz. compounds 28 and 29, were obtained in yields of 52% and 21%, respectively. The lack of any products of intramolecular enolate alkylation processes may be attributed to the limited nucleophilicity and/or the ready protonation of the intermediate samarium enolate. $25,26$

Sufficient quantities of compound 28 were obtained by the means just described to allow for an investigation of the intramolecular enolate alkylation reaction under more conventional conditions. Thus, treatment of a THF solution of ketone 28 maintained at -78 °C with 1.2 mol equiv of lithium hexamethyldisilazide (LiHMDS) (Scheme 4) and then allowing the reaction mixture to warm to 18 °C provided, after work up and chromatographic

purification, compound 32 in 53% yield. All the spectroscopic data obtained on this material were in full accord with the assigned structure but final confirmation of this was secured by a singlecrystal X-ray analysis. The derived ORTEP is shown in [Figure 2](#page-4-0) while other details of this analysis are presented in the [Experimental](#page-4-0) [section](#page-4-0). The formation of this product must arise through selective formation of precursor enolate 30, which then engages in intramolecular alkylation with the tosyloxy-bearing side-chain to give ketone 32 incorporating the tricyclic framework of 2 isocyanoallopupukeanane but carrying the methyl group in the C6a rather than the desired C6 position. Interestingly, no evidence could be obtained for the formation of the isomeric ketone 33 that would arise from intramolecular alkylation of intermediate 31.

Figure 2. Molecular structure of compound 32 ($C_{18}H_{20}O_3$). Anisotropic displacement ellipsoids show 30% probability levels. Hydrogen atoms are drawn as circles with small radii.

Presumably, the selective formation of enolate 30 over isomer **31** under the conditions of thermodynamic control^{[27](#page-11-0)} defined above is a reflection of the reduction in torsional strain (between the angular methyl and the syn-1,2-related methylene proton adjacent to the carbonyl group) associated with the conversion $28\rightarrow 30$. This reduction is greater than that which would be encountered in the equivalent process leading to 31 (where the corresponding reduction in torsional strain would 'only' be that arising from loss of the interaction between the angular hydrogen and the syn-1,2-related methylene proton adjacent to the carbonyl group). In principle, carrying out enolate formation under conditions of kinetic control (viz. adding the substrate ketone 28 to a solution of the base) 27 might be expected to lead to enolate 31 and thence the tricyclic product 33, which bears a pseudo-enantiomeric relationship to isomer 32. Unfortunately, a lack of sufficient quantities of ketone 28 has prevented us from conducting the relevant experiments. Current efforts are directed towards achieving a much more efficient route to compound 28 and the results of these will be reported in due course.

2.4. Potentially enantiodivergent routes to the octahydro-1,6 methano-1H-indene framework associated with 2 isocyanoallopupukeanane

The lack of certainty regarding the absolute configuration of the naturally occurring form of 2-isocyanoallopupukeanane^{[6](#page-11-0)} has prompted us to consider ways in which either enantiomeric form of this compound could be synthesised using the now validated strategy shown in [Figure 1.](#page-1-0) Three distinct ways of achieving this seem possible. First of all, the enantiomeric form of the starting material, viz. compound ent-8, is available from p-iodotoluene using methodology reported by Boyd et al.^{[28](#page-11-0)} Accordingly, the optical antipodes of all of compounds $9-29$ and 32 are automatically available using the chemistries reported herein. Another possible mode of entry into the other enantiomeric series would involve reversing the facial selectively of the initial Diels-Alder cycloaddition reaction involving cis-1,2-dihydrocatechol 8 and/or its derivatives since an α -face addition process affords the pseudoenantiomeric form of the compound (e.g., ketone 10) arising from the corresponding β -face addition reaction. In recent work^{10a,b,29} we have shown that such facial selectivities can be controlled, to some extent at least, by using either compound 8 or a protected form thereof (e.g., acetonide **9**) as the 4π -addend in the cycloaddition process. A third possible mode of enantiomeric 'switching' arises from the pseudo-symmetrical nature of ketonic systems such as compound 28. In particular, if the non-methylated variant of this diquinane could be obtained from the known, benzene-derived cis -1,2-dihydrocatechol¹³ then this could be desymmetrised using the relevant Koga-Simpkins type-base^{[30](#page-11-0)} to generate either enantiomeric form of the corresponding enolate and thence, through intramolecular alkylation, either enantiomeric form of the title framework.³¹ The required methyl group could then be introduced through a conventional dehydrogenation/conjugate addition reaction sequence. Efforts to examine all of these possibilities are now underway in our laboratories. Results will be reported in due course.

3. Experimental section

3.1. General experimental procedures

Proton (1 H) and carbon (13 C) NMR spectra were recorded on a Varian Gemini machine operating at 300 or 75 MHz, respectively. Unless otherwise specified, spectra were acquired at 20 \degree C in deuterochloroform $(CDCI₃)$ that had been stored over anhydrous sodium carbonate. Chemical shifts are recorded as δ values in parts per million (ppm). Infrared spectra (v_{max}) were normally recorded on a Perkin-Elmer 1800 Series FTIR Spectrometer and samples were analysed as thin films on KBr plates (for liquids) or as a KBr disk (for solids). Low-resolution ESI mass spectra were recorded in positiveion mode on a Micromass-Waters LC-ZMD single quadrupole liquid chromatograph-mass spectrometer while low- and high-resolution EImass spectrawere recorded on a Fisons VGAUTOSPEC instrument. Melting points were measured on a Stanford Research Systems Optimelt-Automated Melting Point System and are uncorrected. Optical rotations were measured at the sodium-D line (λ =589 nm) between 17 and 20 °C and at the concentrations (c, in $g/100$ mL) indicated using spectroscopic grade chloroform $(CHCl₃)$ as solvent. Analytical thin layer chromatography (TLC) was performed on aluminium-backed 0.2 mm thick silica gel 60 $F₂₅₄$ plates as supplied by Merck. Eluted plates were visualised using a 254 nm UV lamp and/or by treatment with a suitable dip followed by heating. These dips included a mixture of vanillin/sulfuric acid/ethanol (1 g:1 g:18 mL) or phosphomolybdic acid/ceric sulfate/sulfuric acid (concd)/water (37.5 g: 7.5 g: 37.5 g: 720 mL). The retardation factor (R_f) values cited here have been rounded to the first decimal point. Flash chromatographic separations were carried out following protocols defined by Still et al.^{[32](#page-11-0)} with silica gel 60 (0.040–0.0063 mm) as the stationary phase and using the AR- or HPLC-grade solvents indicated. Starting materials and reagents were generally available from the Sigma-Aldrich, Merck, TCI, Strem or Lancaster Chemical Companies and were either used as supplied or, in the case of liquids, distilled when required. Drying agents and other inorganic salts were purchased from the AJAX, BDH or Unilab Chemical Companies. THF, dichloromethane, acetonitrile and benzene were dried using a Glass Contour solvent purification system that is based upon a technology originally described by Grubbs et al.^{[33](#page-11-0)} Spectroscopic grade solvents were used for all analyses. Where necessary, reactions were performed under a nitrogen or argon atmosphere.

3.2. Specific chemical transformations

3.2.1. Compound 11. LiHMDS (28.8 mL of a 1 M solution in THF, 28.8 mmol, 2 mol equiv) was diluted with diethyl ether (132 mL) then cooled to -78 °C. A solution of ketone 10^{12} 10^{12} 10^{12} (3.00 g, 14.40 mmol) in diethyl ether (12 mL) was then added to the reaction mixture via syringe pump at the rate of 15 mL/h. After addition was complete, the resulting mixture was stirred at -78 °C for 2.5 h then methyl cyanoformate (Mander's reagent) (2.45 g or 2.30 mL, 28.8 mmol, 2 mol equiv) was added via syringe pump at 0.45 mL/h. The ensuing mixture was stirred at -78 $^{\circ}$ C for 0.5 h after which it was poured into dichloromethane/water (300 mL of a 1:1 v/v mixture). The separated aqueous phase was extracted with dichloromethane $(3\times100 \text{ mL})$ and the combined organic layers were washed with brine (1×150 mL) then dried (MgSO₄), filtered and concentrated under reduced pressure. The orange oil thus obtained was subjected to column chromatography (silica, 1:4 v/v ethyl acetate/hexane elution) and concentration of the appropriate fractions (R_f =0.2) gave β -ketoester 11 (2.56 g, 67%) as a white, crystalline solid. This material was used in the subsequent steps of the synthesis (see below). For the purposes of analysis a sample of this material was recrystallised (ethyl acetate/hexane) to give colourless crystals, mp=115-116 °C, $[\alpha]_D$ +210.0 (c 1, CHCl₃). Found: M⁺*, 266.1163; C, 63.43; H, 6.91. C₁₄H₁₈O₅ requires M⁺⁺, 266.1154; C, 63.15; H, 6.81%. ¹H NMR (300 MHz, CDCl₃) δ 6.44 (br dd, J=8.1 and 6.4 Hz, 1H), 5.71 (br dt, $I=8.1$ and 1.5 Hz, 1H), 4.49 (br dd, $I=7.1$ and 3.4 Hz, 1H), 4.08 (dd, $J=7.1$ and 1.5 Hz, 1H), 3.71 (s, 3H), 3.46-3.41 (complex m, 1H), 2.92 (d, $J=1.6$ Hz, 1H), 1.37 (s, 3H), 1.35 (s, 3H), 1.30 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 202.0 (C), 167.8 (C), 133.0 (CH), 129.2 (CH), 110.9 (C), 79.4 (CH), 78.1 (CH), 54.6 (C), 52.8 (CH₃), 50.4 (CH), 38.3 (CH), 25.3 (CH₃), 25.0 (CH₃), 14.6 (CH₃); IR ν_{max} (KBr) 2979, 2949, 2937, 2891, 1747, 1725, 1374, 1262, 1243, 1207, 1163, 1088, 1063, 1034, 973, 714 cm⁻¹; MS (EI, 70 eV) m/z 266 (M⁺⁺, 48%), 251 [(M–CH₃•)⁺, 45], 235 (24), 176 (100), 148 (93), 121 (71), 108 (69), 100 (89), 91 (63), 85 (65), 43 (68).

3.2.2. Compound 12. Method A: A solution of β -ketoester 11 (101 mg, 380 μ mol) in ethanol (17.4 mL) was cooled to 0 °C then treated, in one portion, with sodium borohydride (14.4 mg, 380 µmol, 1 mol equiv). The ensuing mixture was stirred at 0 $^{\circ}$ C for 0.5 h then allowed to warm to 18 °C. After 1 h at this temperature the reaction mixture was treated with ammonium chloride (5 mL of a saturated aqueous solution) and, after a further 5 min, with distilled water (50 mL) then concentrated under reduced pressure to ca. 1/3 of its original volume. The residue so obtained was diluted with dichloromethane (10 mL). The separated aqueous phase was extracted with dichloromethane $(3\times10$ mL) and the combined organic phases were then washed with brine (1×10 mL) before being dried (magnesium sulfate), filtered and concentrated under reduced pressure. The resulting yellow oil was subjected to column chromatography (silica, 1:7:12 v/v/v MeOH/ethyl acetate/hexane elution) to afford two fractions, A and B.

Concentration of fraction A $\left[\frac{R_f}{0.4(4)}\right]$ in 1:7:12 v/v/v MeOH/ ethyl acetate/hexane] yielded an inseparable 1:1.7 mixture of compounds 12 and C8,C9-di-epi-12 (25 mg, 24%) as a clear, colourless oil. Found: M⁺*, 268.1306. C₁₄H₂₀O₅ requires M⁺*, 268.1311. ¹H NMR (300 MHz, CDCl₃) δ (compound **12**) spectrum identical with that derived from a pure sample (see below); ¹H NMR (300 MHz, CDCl₃) δ (compound C8,C9-di-epi-12) 6.03 (dd, J=8.1 and 6.0 Hz, 1H), 5.78–5.70 (complex m, 1H), 4.40 (ddd, $J=7.3$, 3.2 and 0.8 Hz, 1H), 4.35 (dd, J=7.3 and 0.8 Hz, 1H), 3.79 (br d, J=4.4 Hz, 1H), 3.69 (s, 3H), 3.22-3.14 (complex m, 1H), 2.28 (dd, J=4.4 and 1.8 Hz, 1H), 1.36 (s, 3H), 1.32 (s, 3H), 1.28 (s, 3H) (signal due to OH proton obscured or overlapping); ¹³C NMR (75 MHz, CDCl₃) δ (compound **12**) spectrum identical with that derived from a pure sample (see below); 13 C NMR (75 MHz, CDCl₃) δ (compound C8,C9-di-epi-12) 173.7 (C), 134.9 (CH), 129.7 (CH), 108.5 (C), 78.6 (CH), 77.7 (CH), 75.0 (CH), 52.3 (CH₃), 49.9 (CH), 43.9 (C), 37.5 (CH), 25.4 (CH₃), 24.9 (CH_3) , 17.7 (CH_3) ; MS $(EI, 70 \text{ eV})$ m/z 268 $(M^+, 2\%)$, 253 $[(M-CH_3^*)^+]$ 54], 133 (100), 109 (73), 108 (91), 105 (90), 100 (86), 43 (85).

Concentration of fraction B $[R_f=0.4(1)$ in 1:7:12 v/v/v MeOH/ ethyl acetate/hexane] gave compound C9-epi-12 (56 mg, 58%) as a white, crystalline solid, mp=84-86 °C, $[\alpha]_D$ +26.5 (c 0.9, CHCl₃). Found: $(M-CH_3^{\bullet})^+$, 253.1076. C₁₄H₂₀O₅ requires $(M-CH_3^{\bullet})^+$ 253.1076. ¹H NMR (300 MHz, CDCl₃) δ 6.35–6.29 (m, 1H), 5.74 (dd, $J=8.2$ and 1.2 Hz, 1H), 4.17 (dd, $J=7.2$ and 3.6 Hz, 1H), 3.83 (dd, $J=7.2$ and 1.2 Hz, 1H), 3.72 (d, $I=8.6$ Hz, 1H), 3.67 (s, 3H), 3.11-3.06 (complex m, 1H), 2.75 (d, J=8.6 Hz, 1H), 2.10 (br s, 1H), 1.37 (s, 3H), 1.31 (s, 3H), 1.26 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.0 (C), 131.9 (CH) , 131.5 (CH), 109.6 (C), 80.3 (CH), 78.3 (CH), 71.8 (CH), 51.9 (CH₃), 47.8 (CH), 44.9 (C), 36.2 (CH), 25.3 (CH₃), 24.9 (CH₃), 18.2 (CH₃); IR v_{max} (KBr) 3481, 2977, 2934, 2876, 1737, 1454, 1437, 1371, 1346, 1260, 1203, 1171, 1080, 1056, 1031, 883, 836, 731 cm⁻¹; MS (EI, 70 eV) m/z $268 \, (M^+,\langle 1\% \rangle, 253 \, [(M{-}CH_3^{\bullet})^+, 26], 133 \, (63), 109 \, (61), 108 \, (100),$ 105 (47), 80 (43), 43 (40).

Method B (epimerisation of compound C9-epi- 12): DBU (58 µL, 220 μ mol) was added to a solution of compound C9-epi-12 (59 mg, 220μ mol) in benzene (1.1 mL) and the reaction mixture heated at 72 $\,^{\circ}$ C for 21 h then cooled to 18 $\,^{\circ}$ C and diluted with dichloromethane (10 mL). The solution thus obtained was washed with HCl $(2\times10$ mL of a 2 M aqueous solution), sodium bicarbonate $(1\times10$ mL of a saturated aqueous solution) and brine $(1\times10 \text{ mL})$ before being dried, filtered and concentrated under reduced pressure. The resulting lightyellow solid (52 mg), which was comprised of a 1:3.4 mixture of β hydroxyesters C9-epi-**12** and **12** (as determined by 1 H NMR analysis), was subjected to column chromatography (silica,1:7:12 v/v/v MeOH/ ethyl acetate/hexane elution) to afford two fractions, A and B.

Concentration of fraction A $[R_f=0.4(4)$ in 1:7:12 v/v/v MeOH/ethyl acetate/hexane] yielded compound 12 (29 mg, 67% at 73% conversion) as a white, crystalline solid, mp=132.2–132.4 °C, $[\alpha]_D + 47.9$ (c 1, CHCl₃). Found: M^{+•}, 268.1312; C, 62.69; H, 7.54. C₁₄H₂₀O₅ requires M⁺, 268.1311; C, 62.67; H, 7.51%. ¹H NMR (300 MHz, CDCl₃) δ 6.28 $(ddd, J=8.1, 6.7 and 0.8 Hz, 1H), 5.77 (dd, J=8.1 and 1.2 Hz, 1H), 4.23$ $(\text{ddd}, \text{J} = 7.3, 3.4 \text{ and } 1.2 \text{ Hz}, 1\text{H})$, 3.91 $(\text{dd}, \text{J} = 3.3 \text{ and } 1.2 \text{ Hz}, 1\text{H})$, 3.87 (dd, $J=7.4$ and 1.2 Hz, 1H), 3.74 (s, 3H), 3.20-3.15 (complex m, 1H), 2.33 (t, J=3.4 Hz, 1H), 1.59 (s, 1H), 1.37 (s, 3H), 1.31 (s, 3H), 1.25 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 173.3 (C), 133.4 (CH), 131.2 (CH), 109.5 (C), 80.6 (CH), 75.7 (CH), 73.0 (CH), 52.4 (CH), 52.3 (CH₃), 44.9 (C), 36.4 (CH), 25.4 (CH₃), 24.9 (CH₃), 17.8 (CH₃); IR ν_{max} (KBr) 3461, 2976, 2927, 2876, 1731, 1374, 1266, 1248, 1206, 1163, 1078, 1066, 1029, 882, 730 cm⁻¹; MS (EI, 70 eV) m/z 268 (M⁺⁺, 8%), 253 [(M-CH₃⁺)⁺, 49], 133 (100), 109 (64), 108 (63), 105 (69), 100 (67), 43 (71).

Concentration of fraction B $[R_f=0.4(1)$ in 1:7:12 v/v/v MeOH/ ethyl acetate/hexane] afforded β -hydroxyester C9-epi-12 (16 mg, 27% recovery) as a white, crystalline solid that was identical, in all respects, with an authentic sample.

3.2.3. Compound 14 . Step i: Triethylamine (2.27 g, 22.44 mmol, 1.5 mol equiv) was added to a magnetically stirred solution of a ca. 4:1 mixture^{[34](#page-11-0)} of alcohols **12** and C8,C9-di-epi-12 (4.01 g) , 14.96 mmol) in dichloromethane (75 mL) and the resulting mixture cooled to 0° C then treated, dropwise over 5 min, with methanesulfonyl chloride (1.89 g, 16.46 mmol, 1.1 mol equiv). The ensuing mixture was stirred at 0° C for 1 h then at 18 $^{\circ}$ C for 4 h after which it was diluted with dichloromethane (25 mL). The resulting solution was washed with ice-cold water $(1\times100$ mL), hydrochloric acid (1×100 mL of a 2 M aqueous solution), sodium hydrogen carbonate $(1\times100 \text{ mL of a saturated aqueous solution})$ and brine $(1\times100 \text{ mL})$ before being dried (magnesium sulfate), filtered and concentrated under reduced pressure to give an off-white solid

(4.89 g). This material, which was comprised (as determined by $^1\mathrm{H}$ NMR analysis) of a ca. 4:1 mixture of the *mesylates of alcohols* 12 (viz. 13) and C8,C9-di-epi-12, was used directly in the step ii of the reaction sequence.

Step ii: DBU (2.40 mL, 16.07 mmol, 2.6 mol equiv) was added to a solution of the above-mentioned mixture of mesylates (2.14 g, 6.18 mmol) in benzene (30 mL). The resulting mixture was heated to 72 \degree C for 15 h then cooled to 18 \degree C and diluted with dichloromethane (20 mL). The ensuing solution was washed with hydrochloric acid (1×50 mL of a 2 M aqueous solution), sodium hydrogen carbonate (1×50 mL of a saturated aqueous solution) and brine $(1\times50$ mL) then dried (magnesium sulfate), filtered and concentrated under reduced pressure. The yellow oil (1.62 g) thus obtained was subjected to column chromatography (silica, 1:7:12 v/v/v MeOH/ethyl acetate/hexane elution) and concentration of the appropriate fractions (R_f =0.8) then gave compound **14** (1.42 g, 92%) as a light-yellow oil, $[\alpha]_{\text{D}}$ +43.7 (c 1, CHCl₃). Found: $(\text{M--CH}_{3}^{\bullet})^{+}$, 235.0968. C₁₄H₁₈O₄ requires $(M-CH_3^{\bullet})^+$, 235.0970. ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$ δ 6.92 (s, 1H), 6.34–6.28 (complex m, 1H), 5.99-5.95 (complex m, 1H), 4.30-4.25 (complex m, 2H), 3.93-3.89 (complex m, 1H), 3.71 (s, 3H), 1.57 (s, 3H), 1.30 (s, 3H), 1.22 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 164.8 (C), 150.1 (CH), 138.1 (C), 136.2 (CH), 132.1 (CH), 113.2 (C), 82.9 (CH), 79.9 (CH), 51.7 (CH), 47.7 (C), 41.7 (CH₃), 25.8 (CH₃), 25.5 (CH₃), 19.0 (CH₃); IR ν_{max} (KBr) 2977, 2949, 2934, 2905, 1718, 1456, 1437, 1380, 1371, 1263, 1242, 1211, 1162, 1058, 1037, 881, 755, 744, 717 cm⁻¹; MS (EI, 70 eV) m/z 235 [(M-CH₃·)⁺, 33%], 163 (90), 119 (75), 100 (96), 91 (64), 85 (100), 43 (85).

3.2.4. Compound 15. Method A: A solution of α , β -unsaturated ester 14 (897 mg, 3.59 mmol) in THF/MeOH (124 mL of a 7:1 v/v mixture) was cooled to 0 $^{\circ}$ C and sodium borohydride (489 mg, 12.93 mmol, 3.6 mol equiv) was then added in one portion. The ensuing mixture was stirred at 0 $^{\circ}$ C to 18 $^{\circ}$ C for 4.5 h then re-cooled to 0 $^{\circ}$ C and quenched with ammonium chloride (100 mL of a saturated aqueous solution). The resulting mixture was extracted with dichloromethane $(3\times50 \text{ mL})$ and the combined organic extracts were washed with brine $(1\times100 \text{ mL})$ before being dried (magnesium sulfate), filtered and concentrated under reduced pressure. The ensuing light-yellow oil was subjected to column chromatography (silica, 1:7:12 v/v/v MeOH/ethyl acetate/hexane elution) to afford two fractions, A and B.

Concentration of fraction A $[R_f=0.8(4)$ in 1:7:12 v/v/v MeOH/ethyl acetate/hexane] gave compound 15 (333 mg, 37%) as a clear, colourless oil, [α] $_{\rm D}$ – 1.6 (c 1, CHCl₃). Found: M⁺⁺, 252.1357. C₁₄H₂₀O₄ requires M⁺⁺, 252.1362. ¹H NMR (300 MHz, CDCl₃) δ 6.11 (ddd, J=8.1, 6.3 and 0.9 Hz, 1H), 5.83 (dd, J=8.1 and 0.9 Hz, 1H), 4.22 (ddd, J=7.2, 3.3 and 0.9 Hz, 1H), 3.88 (dd, J=7.2 and 1.2 Hz, 1H), 3.69 (s, 3H), 3.16-3.10 (complex m, 1H), 2.47 (ddd, J=11.4, 5.4 and 3.0 Hz, 1H), 1.65 (dd, J=13.5 and 5.4 Hz, 1H), 1.31 (dd, J=13.5 and 11.4 Hz, 1H), 1.29 (s, 3H), 1.25 (s, 3H), 1.23 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 174.8 (C), 136.7 (CH), 130.0 (CH) , 107.9 (C), 82.6 (CH), 76.1 (CH), 52.0 (CH₃), 40.8 (CH), 38.5 (C), 37.3 (CH), 31.4 (CH₂), 25.4 (CH₃), 24.9 (CH₃), 21.5 (CH₃); IR ν_{max} (KBr) 2976, 2955, 2936, 2900, 2873,1733,1458,1435,1374,1343,1297,1265,1239, 1205, 1165, 1070, 1055, 1021, 997, 885, 724 cm⁻¹; MS (EI, 70 eV) m/z $252 (M^+$, 13%), 237 $[(M-CH_3^{\bullet})^+, 44]$, 221 $[(M-CH_3O^{\bullet})^+, 16]$, 194 (66), 135 (100), 134 (60), 117 (55), 105 (55), 91 (64).

Concentration of fraction B $\left[Re=0.7(9)$ in 1:7:12 v/v/v MeOH/ethyl acetate/hexane] yielded compound C9-epi-15 (398 mg, 44%) as a clear, colourless oil, [α] $_{\rm D}$ –26.2 (c 1, CHCl $_3$). Found: M $^+$, 252.1363. $C_{14}H_{20}O_4$ requires M⁺, 252.1362. ¹H NMR (300 MHz, CDCl₃) δ 5.96 $(dd, J=8.1$ and 6.3 Hz, 1H), 5.88 (dd, J=8.1 and 0.6 Hz, 1H), 4.24 (ddd, $J=7.2$, 3.3 and 0.9 Hz, 1H), 3.84 (dd, $J=7.2$ and 1.2 Hz, 1H), 3.65 (s, 3H), 3.23–3.18 (complex m, 1H), 2.50 (ddd, $J=10.2$, 5.1 and 2.4 Hz, 1H), 1.59 (dd, J=13.5 and 5.1 Hz, 1H), 1.39 (dd, J=13.5 and 10.2 Hz, 1H), 1.30 (s, 3H), 1.26 (s, 3H), 1.26 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 174.6 (C) , 136.9 (CH), 127.8 (CH), 108.6 (C), 82.7 (CH), 78.8 (CH), 52.0 (CH₃), 39.6 (CH), 38.1 (C), 37.5 (CH), 32.3 (CH2), 25.5 (CH3), 25.0 (CH3), 21.5 (CH₃); IR v_{max} (KBr) 2971, 2956, 2931, 2873, 1738, 1373, 1285, 1254, 1203, 1167, 1063, 885, 714 cm⁻¹; MS (EI, 70 eV) m/z 252 (M⁺⁻, 7%), 237 [(M–CH₃·)⁺, 61], 221 [(M–CH₃O·)⁺, 27], 194 (84), 162 (84), 135 (100), 134 (83), 117 (70), 105 (70), 100 (81), 93 (70).

Method B (epimerisation of compound C9-epi-15): A solution of ester C9-epi-15 (368 mg, 1.46 mmol) in MeOH (27 mL) was cooled to 0 °C then sodium methoxide [generated from NaH (105 mg, 4.38 mmol, 3 mol equiv) and MeOH (18 mL)] was added. After 5 min, the cooling bath was removed and the reaction mixture heated to 70 \degree C for 24 h then allowed to cool to 18 \degree C and quenched with ammonium chloride (20 mL of a saturated aqueous solution). The separated aqueous phase was extracted with dichloromethane $(3\times20 \text{ mL})$ then the combined organic phases were washed with brine $(1\times10 \text{ mL})$ before being dried (magnesium sulfate), filtered and concentrated under reduced pressure to yield a clear, colourless oil. Subjection of this material to column chromatography (silica, 1:7:12 v/v/v MeOH/ethyl acetate/hexane elution) afforded two fractions, A and B.

Concentration of fraction A $[R_f=0.8(4)$ in 1:7:12 v/v/v MeOH/ ethyl acetate/hexane] gave ester 15 (62 mg, 36% at 47% conversion) as a clear, colourless oil that was identical, in all respects, with an authentic sample.

Concentration of fraction B $[R_f=0.7(9)$ in 1:7:12 v/v/v MeOH/ ethyl acetate/hexane] afforded ester C9-epi-15 (196 mg, 53% recovery) as a clear, colourless oil that was identical, in all respects, with an authentic sample.

3.2.5. Compounds 16 and 17. Method A (reduction of ester 15): A magnetically stirred solution of ester **15** (50 mg, 198 μ mol) in dichloromethane (5.20 mL) was cooled to -78 °C then DIBAL $(0.34 \text{ mL of a 1 M solution in hexane, } 340 \text{ µmol, } 1.7 \text{ mol equity})$ was added dropwise over 10 min. The resulting mixture was stirred at -78 °C for 1.25 h then quenched with potassium sodium tartrate (2 mL of a saturated aqueous solution), warmed to 18 $^{\circ}$ C and stirred at this temperature for 15 h. The separated aqueous layer was extracted with dichloromethane $(3\times2$ mL) and the combined organic phases were then dried (magnesium sulfate), filtered and concentrated under reduced pressure to yield a clear, colourless oil (46 mg). Subjection of this material to column chromatography (silica, 1:4 v/v ethyl acetate/hexane elution) afforded two fractions, A and B.

Concentration of fraction A (R_f =0.4 in 1:4 v/v ethyl acetate/hexane) gave aldehyde 17 (13 mg, 30%) as a white, crystalline solid, $mp = 59 - 62 °C$, $[\alpha]_D - 31.0$ (c 1, CHCl₃). Found: M⁺⁺, 222.1253; C, 70.05; H, 8.00. C $_{13}$ H $_{18}$ O $_{3}$ requires M⁺*, 222.1256; C, 70.25; H, 8.16%. ¹H NMR $(500$ MHz, CDCl₃) δ 9.77 (s, 1H), 6.16 (dd, J=8.1 and 6.8 Hz, 1H), 5.88 (d, $J=8.1$ Hz, 1H), 4.05 (ddd, J=7.2, 3.2 and 1.0 Hz, 1H), 3.78 (dd, J=7.2 and 1.0 Hz, 1H), 3.29-3.27 (complex m, 1H), 2.47 (ddd, J=11.2, 5.4 and 2.9 Hz, 1H), 1.70 (dd, J=13.6 and 5.4 Hz, 1H), 1.29 (s, 3H), 1.27 (s, 3H), 1.21 (s, 3H), 1.20 (dd, J=13.6 and 11.2 Hz, 1H); ¹³C NMR (125 MHz, CDCl3) d 202.4 (CH), 137.5 (CH), 129.5 (CH), 108.1 (C), 82.5 (CH), 75.9 (CH) , 49.5 (CH), 38.8 (C), 35.4 (CH), 28.1 (CH₂), 25.4 (CH₃), 24.9 (CH₃), 21.6 (CH₃); IR v_{max} (KBr) 2975, 2955, 2934, 2872, 1722, 1458, 1373, $1264, 1254, 1208, 1165, 1135, 1071, 1058, 971, 884, 824, 729, 699$ cm⁻¹; MS (EI, 70 eV) m/z 223 [(M+H)⁺, 15%], 222 (M⁺, 5), 207 [(M–CH₃•)⁺, 46], 164 (63), 135 (98), 117 (65), 93 (67), 92 (100), 91 (63), 43 (75).

Concentration of fraction B (R_f =0.1 in 1:4 v/v ethyl acetate/ hexane) afforded *alcohol* **16** (25 mg, 57%) as a clear, colourless oil, $[\alpha]_{D}$ –17.3 (c 0.9, CHCl₃). Found: $(M-CH_{3})^{+}$, 209.1179. C₁₃H₂₀O₃ requires (M–CH₃•)⁺, 209.1178. ¹H NMR (300 MHz, CDCl₃) δ 6.16 (dd, $J=8.2$ and 6.9 Hz, 1H), 5.80 (d, $J=8.2$ Hz, 1H), 4.39 (dd, $J=7.3$ and 3.2 Hz, 1H), 3.79 (d, $J=7.3$ Hz, 1H), 3.64-3.47 (complex m, 2H), 2.96-2.92 (complex m, 1H), 1.85-1.74 (complex m, 2H), 1.35 (dd, $J=13.3$ and 11.1 Hz, 1H), 1.31 (s, 3H), 1.26 (s, 3H), 1.22 (s, 3H), 0.69 $(dd, J=13.3$ and 5.9 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 135.6 (CH), 131.9 (CH), 107.7 (C), 83.1 (CH), 75.6 (CH), 65.0 (CH₂), 38.6 (CH), 38.4 (C), 35.9 (CH), 33.1 (CH₂), 25.5 (CH₃), 24.9 (CH₃), 21.7 (CH₃); IR ν_{max} (KBr) 3417, 3044, 2970, 2933, 2869, 1457, 1374, 1269, 1246, 1207, 1165, 1075, 1058, 1028, 885, 730, 705, 691, 513 cm⁻¹; MS (EI, 70 eV) m/z 209 [(M–CH₃•)⁺, 25%], 166 (60), 135 (100), 93 (63).

Method B (oxidation of alcohol **16**): A magnetically stirred solution of alcohol 16 (980 mg, 4.37 mmol) in dichloromethane/DMSO (62 mL of a 1:1 v/v mixture) was cooled to 0 $^{\circ}$ C then treated with triethylamine (3 mL, 21.85 mmol, 5 mol equiv) and sulfur trioxide pyridine complex (2.09 g, 13.11 mmol, 3 mol equiv). The ensuing mixture was stirred at 0 $\rm{^{\circ}C}$ for 1 h, diluted with diethyl ether (50 mL) then washed with hydrochloric acid (1 \times 10 mL of a 1 M aqueous solution), sodium hydrogen carbonate $(1\times10 \text{ mL of a sat-})$ urated aqueous solution) and brine $(1\times10 \text{ mL})$ before being dried (magnesium sulfate), filtered and concentrated under reduced pressure. The resulting light-orange oil (922 mg) was subjected to column chromatography (silica, 1:5:14 v/v/v MeOH/ethyl acetate/ hexane elution) and gave, after concentration of the appropriate fractions, aldehyde 17 (497 mg, 51%) as a white, crystalline solid that was identical, in all respects, with an authentic sample.

Method C (reduction of ester C9-epi-15 and epimerisation of the resulting aldehyde C9-epi-17). Step i: A magnetically stirred solution of ester C9-epi-15 (100 mg, 0.40 mmol) in dichloromethane (10 mL) was cooled to -78 °C then DIBAL (0.48 mL of a 1 M solution in hexane, 0.48 mmol, 1.2 mol equiv) was added dropwise over 10 min and the ensuing mixture then stirred at -78 °C for 2 min before being quenched with potassium sodium tartrate (5 mL of a saturated solution), warmed to 18 $^{\circ}$ C and stirred at this temperature for 5 h. The separated aqueous phase was extracted with dichloromethane $(3\times10 \text{ mL})$ then the combined organic phases were washed with brine (1×5 mL) then dried (magnesium sulfate), filtered and concentrated under reduced pressure to give a clear, colourless oil. Subjection of this material to column chromatography (silica, $1:4\rightarrow1:1$ v/v ethyl acetate/hexane gradient elution) afforded two fractions, A and B.

Concentration of fraction A (R_f =0.4 in 1:4 v/v ethyl acetate/hexane) gave aldehyde 17 (4 mg, 5%) as a white, crystalline solid. This material was identical, in all respects, with an authentic sample.

Concentration of fraction B (R_f =0.3 in 1:4 v/v ethyl acetate/ hexane) afforded compound C9-epi-17 (58 mg, 65%) as a clear, colourless oil, [α] $_D + 15.3$ (c 1.15, CHCl3). Found: (M–CH3 $^{\bullet}$)⁺, 207.1026. $C_{13}H_{18}O_3$ requires (M–CH₃·)⁺, 207.1021. ¹H NMR (300 MHz, CDCl₃) δ 9.49 (d, J=1.1 Hz, 1H), 5.95-5.86 (complex m, 2H), 4.32 (ddd, $J=7.2$, 3.3 and 1.0 Hz, 1H), 3.89 (dd, $J=7.2$ and 1.0 Hz, 1H), 3.25 -3.20 (complex m, 1H), 2.44 (dddd, J=9.9, 4.7, 2.2 and 1.0 Hz, 1H), 1.64 (dd, $J=13.6$ and 4.7 Hz, 1H), 1.31 (dd, $J=13.6$ and 9.9 Hz, 1H), 1.30 (s, 3H), 1.28 (s, 3H), 1.27 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 201.9 (CH), 137.9 (CH), 127.1 (CH), 108.7 (C), 83.1 (CH), 79.0 (CH), 48.0 (CH), 38.4 (C), 35.9 (CH), 29.6 (CH₂), 25.4 (CH₃), 25.0 (CH₃), 21.5 (CH₃); IR ν_{max} (KBr) 2975, 2954, 2932, 2873, 1726, 1458, 1374, 1277, 1251, 1209, 1166, 1121, 1068, 1023, 884, 728 cm⁻¹; MS (EI, 70 eV) m/z 223 $[(M+H)^{+}$, 20%], 222 (M⁺⁺, 1), 207 $[(M-CH₃[*])⁺$, 16], 135 (79), 117 (52), 100 (51), 93 (49), 91 (63), 85 (46), 43 (100).

Step ii: DBU (26 μ L, 171 μ mol, 1 mol equiv) was added to a magnetically stirred solution of aldehyde C9-epi-17 (38 mg, 171 μ mol) in benzene (1 mL) and the resulting mixture heated at 70 °C for 16 h, then cooled to 18 $^{\circ}$ C and diluted with dichloromethane (4 mL) before being washed successively with hydrochloric acid $(1\times2 \text{ mL of a 2 M})$ aqueous solution), sodium hydrogen carbonate $(1\times2 \text{ mL of a satu-})$ rated aqueous solution) and brine $(1\times2$ mL) then dried (magnesium sulfate), filtered and concentrated under reduced pressure. ¹H NMR analysis of the resulting yellow oil (33 mg) established that this was comprised of a ca. 1:4.8 mixture of aldehydes C9-epi-17 and 17.

3.2.6. Compound **18**. MePPh₃Br was dried at 100 °C for 15 h then cooled and stored under nitrogen. Some of the dried material (2.05 g, 5.73 mmol, 3.0 mol equiv) was stirred in THF (19.1 mL) then

cooled to 0° C and treated, dropwise over 0.5 h, with NaHMDS (4.58 mL of a 1 M solution in THF, 4.58 mmol, 2.4 mol equiv). The resulting mixture was stirred at 0 \degree C for 2.5 h and the bright-yellow reaction mixture so-formed was treated, dropwise over 0.5 h, with a solution of aldehyde 17 (424 mg, 1.91 mmol) in THF (8.50 mL). Stirring was continued at 0 $^{\circ}$ C for 2 h then the reaction mixture was quenched with ammonium chloride (10 mL of a saturated aqueous solution) and diluted with dichloromethane (10 mL). The separated aqueous phase was extracted with dichloromethane $(3\times20 \text{ mL})$ then the combined organic phases were washed with brine $(1\times10$ mL) before being dried (magnesium sulfate), filtered and concentrated under reduced pressure. The orange oil thus obtained was subjected to column chromatography (silica, 1:9 v/v ethyl acetate/hexane elution) and concentration of the appropriate fractions $(R_f=0.5)$ then gave olefin **18** (253 mg, 60%) as a clear, colourless oil, $[\alpha]_{\text{D}}$ –45.3 (c 0.55, CHCl₃). Found: (M–H•)⁺, 219.1382. $C_{14}H_{20}O_2$ requires $(M-H^{\bullet})^+$, 219.1385. ¹H NMR (300 MHz, CDCl₃) δ 6.17 (ddd, J=7.8, 6.3 and 1.0 Hz, 1H), 5.83 (ddd, J=17.2, 10.3 and 6.9 Hz, 1H), 5.79 (dd, J=7.8 and 1.0 Hz, 1H), 5.11-5.01 (complex m, 2H), 4.36 (ddd, J=7.4, 3.3 and 1.0 Hz, 1H), 3.81 (dd, J=7.4 and 1.0 Hz, 1H), 2.80–2.74 (complex m, 1H), 2.29–2.18 (complex m, 1H), 1.39 $(dd, J=13.4$ and 11.0 Hz, 1H), 1.32 (s, 3H), 1.26 (s, 3H), 1.24 (s, 3H), 1.01 (dd, J=13.4 and 5.8 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 141.0 (CH), 135.4 (CH), 131.9 (CH), 114.6 (CH₂), 107.6 (C), 83.3 (CH), 75.9 (CH), 40.8 (CH), 39.4 (CH), 38.7 (C), 34.6 (CH₂), 25.5 (CH₃), 24.9 (CH₃), 21.8 (CH₃); IR ν_{max} (KBr) 3045, 2976, 2952, 2937, 2903, 2869, 1638, 1457, 1378, 1371, 1262, 1233, 1207, 1165, 1069, 1056, 995, 912, 885, 861, 809, 729, 707 cm⁻¹; MS (EI, 70 eV) m/z 220 (M⁺⁺, 18%), $219 [(M-H•)+38]$, $205 [(M–CH₃•)+11]$, 163 (68), 105 (64), 57 (100), 43 (46).

3.2.7. Compound 19. DOWEX-50 was activated by washing it twice with MeOH, twice with hydrochloric acid (2 M aqueous solution) and, finally, twice with water. The activated resin (269 mg) thus obtained was added to a magnetically stirred solution of acetonide 18 (269 mg, 1.22 mmol) in MeOH/water (6 mL of a 5:1 v/v mixture) and the resulting suspension heated in an oil bath maintained at 110 °C. After 6 days the reaction mixture was cooled, filtered and the DOWEX washed three times with MeOH. The combined filtrates were concentrated under reduced pressure. The resin was also washed twice with dichloromethane and the filtrate added to the concentrated residue, which was then washed with sodium chloride $(1\times10 \text{ mL of a } 1.5 \text{ M}$ aqueous solution). The separated aqueous phase was extracted with dichloromethane $(3\times10 \text{ mL})$ then the combined organic phases were dried (magnesium sulfate), filtered and concentrated under reduced pressure. The orange oil (201 mg) thus obtained was subjected to column chromatography (silica, $1:4\rightarrow1:1$ v/v ethyl acetate/hexane gradient elution) and concentration of the appropriate fractions (R_f =0.5 in 1:1 v/v ethyl acetate/hexane) then gave diol 19 (146 mg, 66%) as a clear, colourless oil, $[\alpha]_{\text{D}}$ –76.1 (c 1, CHCl₃). Found: M⁺⁺, 180.1144. C₁₁H₁₆O₂ requires M⁺, 180.1150. ¹H NMR (300 MHz, CDCl₃) δ 6.32 (ddd, J=8.1, 7.0 and 0.8 Hz, 1H), 5.90 (dd, $J=8.1$ and 0.8 Hz, 1H), 5.82 (ddd, J=17.0, 10.3 and 6.7 Hz, 1H), 5.10-5.01 (complex m, 2H), 4.05 (ddd, $J=7.6$, 2.5 and 0.8 Hz, 1H), 3.44 (dd, $J=7.6$ and 0.8 Hz, 1H), 2.80 (br s, 2H), 2.67-2.62 (complex m, 1H), 2.20-2.09 (complex m, 1H), 1.38 $(dd, J=13.5$ and 11.2 Hz, 1H), 1.23 (s, 3H), 1.06 (dd, J = 13.5 and 6.2 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 140.8 (CH), 136.6 (CH), 133.4 (CH), 114.9 (CH2), 75.6 (CH), 67.9 (CH), 43.8 (CH), 40.2 (C), 40.0 (CH), 35.3 (CH₂), 21.6 (CH₃); IR ν_{max} (KBr) 3374, 2928, 2869, 1637, 1457, 1403, 1372, 1065, 1031, 994, 959, 910, 726, 703, 601 cm⁻¹; MS (EI, 70 eV) m/z 180 (M^{+•}, 5%), 120 (97), 105 (100), 92 (72), 91 (53).

3.2.8. Compound 20. A magnetically stirred solution of diol 19 (146 mg, 0.81 mmol) in dichloromethane (19.50 mL) was cooled to $0 °C$ then p-TsOH \cdot H₂O (339 mg, 1.78 mmol, 2.2 mol equiv) was

added followed by 4-AcNH-TEMPO (380 mg, 1.78 mmol, 2.2 mol equiv) and the mixture thus obtained was warmed to 18 $^\circ$ C. After 17 h at the latter temperature, the reaction mixture was quenched with sodium hydrogen carbonate (10 mL of a saturated aqueous solution) and extracted with dichloromethane $(3\times20 \text{ mL})$. The combined organic extracts were washed with brine $(1\times10 \text{ mL})$ before being dried (magnesium sulfate), filtered and concentrated under reduced pressure. The resulting orange semi-solid (533 mg) was subjected to column chromatography (silica, 1:4 v/v ethyl acetate/hexane elution) and concentration of the appropriate fractions (R_f =0.3) then gave *acyloin* **20** (118 mg, 81%) as a clear, colourless oil, $[\alpha]_D$ +346.5 (c 0.65, CHCl₃). Found: M⁺⁺, 178.0986. $\rm C_{11}H_{14}O_2$ requires M⁺, 178.0994. ¹H NMR (300 MHz, CDCl₃) δ 6.19 (dd, J=7.7 and 6.6 Hz, 1H), 6.11 (d, J=7.7 Hz, 1H), 5.68 (ddd, J=17.0, 10.3 and 7.3 Hz, 1H), 5.10-4.98 (complex m, 2H), 3.37 (s, 1H), 3.20 $(dd, J=6.6$ and 2.2 Hz, 1H), 2.73 (br s, 1H), 2.61–2.50 (complex m, 1H), 1.80 (dd, $J=13.4$ and 11.7 Hz, 1H), 1.43 (dd, $J=13.4$ and 4.7 Hz, 1H), 1.31 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 211.6 (C), 140.7 (CH), 140.5 (CH), 126.9 (CH), 115.1 (CH₂), 75.1 (CH), 52.7 (CH), 42.2 (C), 39.0 (CH), 37.9 (CH₂), 20.0 (CH₃); IR ν_{max} (KBr) 3448, 2970, 2954, 2931, 2869, 1725, 1639, 1457, 1115, 1068, 990, 915, 776, 709 cm $^{-1};$ MS (EI, 70 eV) m/z 178 (M^{+•}, 8%), 106 (72), 105 (100), 91 (71), 79 (70), 43 (73), 39 (64), 32 (67).

3.2.9. Compound 21. A magnetically stirred solution of acyloin 20 (118 mg, 0.66 mmol) in dichloromethane (21 mL) was cooled to 0 $^{\circ}$ C then treated with triethylamine (0.46 mL, 3.30 mmol, 5 mol equiv), DMAP (9 mg, 0.07 mmol, 10 mol %) and benzoyl chloride (0.15 mL, 1.32 mmol, 2 mol equiv). The ensuing mixture was allowed to warm to 18 °C and after 16 h it was quenched with sodium hydrogen carbonate (10 mL of a saturated aqueous solution). The separated aqueous phase was extracted with dichloromethane $(3\times20 \text{ mL})$ then the combined organic extracts were washed with brine $(1\times10$ mL) before being dried (magnesium sulfate), filtered and concentrated under reduced pressure. The yellow semi-solid (301 mg) thus obtained was subjected to column chromatography (silica,1:4 v/v ethyl acetate/hexane elution) and concentration of the appropriate fractions $(R_f=0.6)$ gave keto-ester **21** (199 mg, 0.71 mmol, quant.) as a clear, colourless oil, α _D +255.9 (c 1, CHCl₃). Found: M⁺, 282.1255. C₁₈H₁₈O₃ requires M⁺⁻, 282.1256. ¹H NMR (300 MHz, CDCl₃) δ 8.02-7.98 (complex m, 2H), 7.59-7.52 (complex m, 1H), 7.46–7.39 (complex m, 2H), 6.32 (dd, J =7.8 and 6.7 Hz, 1H), 6.20 (d, J=7.8 Hz, 1H), 5.76 (ddd, J=17.2, 10.3 and 7.4 Hz, 1H), 5.16-5.05 (complex m, 2H), 5.08 (s, 1H), 3.26 (ddd, J=6.7, 2.9 and 1.1 Hz, 1H), 2.67-2.56 (complex m, 1H), 1.89 (dd, $J=13.6$ and 11.5 Hz, 1H), 1.62 (dd, J=13.6 and 4.9 Hz, 1H), 1.22 (s, 3H); ¹³C NMR (75 MHz, CDCl3) d 205.3 (C), 166.1 (C), 140.0 (CH), 139.8 (CH), 133.2 (CH), 129.9 (CH), 129.4 (C), 128.3 (CH), 127.6 (CH), 115.6 (CH₂), 74.5 (CH), 53.5 (CH), 41.3 (C), 39.5 (CH), 37.5 (CH₂), 20.1 (CH₃); IR ν_{max} (KBr) 2965, 2925, 2869, 2853, 1741, 1724, 1451, 1328, 1266, 1254, 1177, 1111, 1070, $1029, 921, 708$ cm⁻¹; MS (EI, 70 eV) m/z 282 (M⁺⁺, 21%), 160 (37), 132 (62), 120 (41), 106 (76), 105 (100), 91 (30), 77 (89), 51 (43).

3.2.10. Compound 22. A magnetically stirred and deoxygenated solution of keto-ester 21 (186 mg, 0.66 mmol) and acetophenone (0.23 mL, 1.98 mmol, 3 mol equiv) in acetone (300 mL) was placed in a quartex immersion well photoreactor (Ace Glass Inc., 500 mL) equipped with a Pyrex filter. The mixture was subjected to irradiation, at 18 °C, with a Hanovia 450 W medium pressure mercuryvapour lamp. After 4.67 h the reaction mixture was removed from the photoreactor and concentrated under reduced pressure to give a yellow oil (459 mg) that was subjected to column chromatography (silica, 1:4 v/v ethyl acetate/hexane elution) and thereby affording two fractions, A and B.

Concentration of fraction A (R_f =0.5) yielded a mixture of acetophenone and the $C4$ - α -form of diquinane 22 (49 mg) as a clear, colourless oil. Resubjection of the material to flash chromatography (silica, 1:9 v/v ethyl acetate/hexane elution) and concentration of the appropriate fractions (R_f =0.2) then gave the C4- α -form of diquinane 22 (29 mg, 15%) as a clear, colourless oil, $[\alpha]_D$ +139.1 (c 0.9, CHCl₃). Found: M^{+•}, 282.1260. C₁₈H₁₈O₃ requires M⁺°, 282.1256. ¹H NMR (300 MHz, CDCl₃) δ 8.06–8.02 (complex m, 2H), 7.60–7.54 (complex m, 1H), $7.47-7.40$ (complex m, 2H), 5.82 (ddd, $J=17.3$, 10.4) and 5.6 Hz, 1H), $5.19-5.02$ (complex m, 2H), 5.08 (br s, 1H), 3.46 -3.35 (complex m, 1H), 2.61 (ddd, $J=6.0$, 5.1 and 1.0 Hz, 1H), 2.43 (dd, $J=13.5$ and 11.0 Hz, 1H), 2.35-2.30 (m, 1H), 2.24-2.22 (m, 1H), 1.89 (dt, $J=13.5$ and 1.4 Hz, 1H), 1.28 (s, 3H); ¹³C NMR (75 MHz, CDCl3) d 210.9 (C), 165.6 (C), 139.5 (CH), 133.2 (CH), 129.9 (CH), 129.5 (C) , 128.4 (CH), 115.8 (CH₂), 83.5 (CH), 49.9 (C), 49.8 (CH₂), 43.0 (CH), 42.0 (CH), 39.8 (CH), 38.4 (CH), 19.3 (CH₃); IR ν_{max} (KBr) 2970, 2935, 2874, 1722, 1451, 1266 (br), 1177, 1107, 1094, 1069, 1026, 990, 957, 915, 852, 709, 668 cm $^{-1}$; MS (EI, 70 eV) m/z 282 (M⁺, 2%), 160 (25), 132 (45), 106 (55), 105 (71), 91 (37), 77 (100).

Concentration of fraction B (R_f =0.5) gave the C4- β -form of diquinane 22 (30 mg, 17%) as a clear, colourless oil, α _D +60.2 (c 0.6, CHCl₃). Found: M^{+•}, 282.1256. C₁₈H₁₈O₃ requires M^{+•}, 282.1256. ¹H NMR (300 MHz, CDCl₃) δ 8.07-8.03 (complex m, 2H), 7.61-7.55 (complex m, 1H), 7.48-7.42 (complex m, 2H), 5.80 (ddd, $J=17.0$, 10.3) and 7.0 Hz, 1H), 5.44 (t, J=1.5 Hz, 1H), 5.11 (dt, J=17.0 and 1.5 Hz, 1H), 5.01 (dt, 10.3 and 1.5 Hz, 1H), 3.45-3.35 (complex m, 1H), 2.37 $(dd, J=5.9$ and 4.8 Hz, 1H), 2.28 (dt, $J=9.8$ and 5.9 Hz, 1H), 2.12 (ddd, $J=14.0$, 11.0 and 1.5 Hz, 1H), 2.05–1.98 (complex m, 2H), 1.45 (s, 3H); 13° C NMR (75 MHz, CDCl₃) δ 207.1 (C), 165.4 (C), 139.2 (CH), 133.3 (CH), 129.9 (CH), 129.3 (C), 128.4 (CH), 115.3 (CH₂), 82.1 (CH), 49.3 (C), 44.2 (CH), 43.5 (CH₂), 36.4 (CH), 35.8 (CH), 32.6 (CH), 24.3 (CH₃); IR v_{max} (KBr) 2959, 1739, 1724, 1452, 1331, 1269, 1248, 1177, 1113, 1097, 1071, 1026, 1000, 916, 850, 709 cm⁻¹; MS (EI, 70 eV) m/z 282 (Mþ , 11%), 160 (37), 132 (49), 106 (52), 105 (100), 77 (69).

3.2.11. Compound 23. A magnetically stirred solution of the $C4-\alpha$ and β -epimeric forms of compound 22 (296 mg, 1.05 mmol) in THF (10.5 mL) containing MeOH (5.3 mL) was cooled to -78 °C then samarium(II) diiodide (23.1 mL of a 0.1 M solution in THF, 2.31 mmol, 2.2 mol equiv) was added dropwise over 0.5 h. Stirring was continued at -78 °C for 5 min then the reaction mixture was quenched with potassium carbonate (20 mL of a saturated aqueous solution) before being allowed to slowly warm to 18 \degree C. The separated aqueous phase was extracted with diethyl ether $(3\times50 \text{ mL})$ and the combined organic phases were washed with brine $(1\times20 \text{ mL})$ then dried (magnesium sulfate), filtered and concentrated under reduced pressure. The yellow oil (289 mg) thus obtained was subjected to column chromatography (silica, 1:39 v/v ethyl acetate/dichloromethane elution) and concentration of the appropriate fractions (R_f =0.5) gave diquinane 23 (93 mg, 54%) as a clear, colourless oil, $[\alpha]_{\text{D}} +$ 74.0 (c 0.6, CHCl₃). Found: M⁺⁺, 162.1044. $C_{11}H_{14}O$ requires M⁺, 162.1045. ¹H NMR (300 MHz, CDCl₃) δ 5.82 (ddd, J=17.2, 10.3 and 5.8 Hz, 1H), 5.08 (dt, J=17.2 and 1.7 Hz, 1H), 4.97 (dt, J 10.3 and 1.7 Hz, 1H), 3.37-3.26 (complex m, 1H), 2.45 (ddd, J=6.3, 4.9 and 0.8 Hz, 1H), 2.33–2.22 (complex m, 2H), 2.20–2.04 (complex m, 2H), 1.96 (m, 1H), 1.63 (d, J=12.6 Hz, 1H), 1.36 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 215.5 (C), 139.9 (CH), 115.0 (CH₂), 55.0 $(CH₂), 52.4 (CH₂), 46.3 (C), 43.4 (CH), 42.3 (CH), 39.6 (CH), 36.8 (CH),$ 25.7 (CH₃); IR v_{max} (KBr) 2954, 2928, 2870, 1724, 1454, 1407, 1331, 1313, 1250, 1197, 1096, 989, 965, 914, 885, 871 cm⁻¹; MS (EI, 70 eV) m/z 162 (M⁺*, 11%), 120 (50), 105 (100), 77 (45).

3.2.12. Compound 24. A magnetically stirred solution of diquinane 23 (52 mg, 321 μ mol) in acetone/water (2 mL of a 1:1 v/v mixture) was cooled to 0° C then *N*-methylmorpholine *N*-oxide (45 mg, 385 µmol, 1.2 mol equiv) and osmium tetroxide (1.22 mL of a 0.1 M solution in tert-butanol, 122μ mol, 0.38 mol equiv) were added. The ensuing mixture was allowed to warm to 18 \degree C and after 4.5 h at

this temperature it was quenched with sodium hydrogensulfite (4 mL of a saturated aqueous solution) then stirred at 18 $^{\circ}$ C for another hour. The resulting mixture was diluted with diethyl ether (4 mL) and just enough water to dissolve any solids. Solid sodium chloride was then added to saturate the aqueous phase which, after separation, was extracted with diethyl ether $(3\times10 \text{ mL})$ then dichloromethane (3×10 mL). The combined organic extracts were dried (magnesium sulfate), filtered and concentrated under reduced pressure. Subjection of the resulting clear, colourless oil to column chromatography (silica, ethyl acetate \rightarrow 1:9 v/v MeOH/ethyl acetate gradient elution) and concentration of the appropriate fractions (R_f =0.4 in 1:9 v/v methanol/ethyl acetate) then gave a 1:1 mixture of the epimeric forms of diol 24 (30 mg, 48%) as a clear, colourless oil. Found: M⁺, 196.1098. C₁₁H₁₆O₃ requires M⁺, 196.1099. ¹H NMR (300 MHz, CDCl₃) δ 3.84 (dd, J=11.3 and 2.2 Hz, 0.5H), 3.66–3.58 (complex m, 0.5H), 3.50 (br s, 2H), 3.45 (dd, $J=11.3$ and 7.4 Hz, 0.5H), $3.40-3.30$ (complex m, 1.5H), $2.71-2.55$ (complex m, 1H), $2.54-2.48$ (complex m, 1H), $2.43-2.30$ (complex m, 1H), 2.27–1.80 (complex m, 5H), 1.37 (s, 1.5H), 1.36 (s, 1.5H); ^{13}C NMR (75 MHz, CDCl₃) δ 217.6 (C), 217.4 (C), 74.2 (CH), 74.1 (CH), 66.2 $(CH₂), 65.4 (CH₂), 56.4 (CH₂), 56.0 (CH₂), 48.8 (CH₂), 48.6 (CH₂), 46.6)$ (C) , 46.2 (C) , 43.8 $(2\times CH)$, 42.9 (CH) , 42.8 (CH) , 39.3 (CH) , 39.1 (CH) , 37.1 (CH), 35.4 (CH), 25.9 (CH₃), 25.8 (CH₃); IR ν_{max} (KBr) 3405, 2951, 2928, 2871, 1709, 1454, 1408, 1379, 1360, 1335, 1312, 1254, 1200, 1161, 1101, 1076, 1041, 964, 920, 879, 813, 735 cm⁻¹; MS (EI, 70 eV) m/z 196 (M⁺⁺, 2%), 178 [(M-H₂O⁺)⁺, 14], 165 (75), 136 (96), 95 (94), 94 (88), 93 (100), 91 (77), 43 (75).

3.2.13. Compound 25. Dibutyltin(IV) oxide (8 mg, 32 μ mol, 20 mol %) then triethylamine (21 μ L, 153 μ mol, 1 mol equiv) were added to a magnetically stirred solution of a 1:1 mixture of the epimeric forms of diol 24 (30 mg, 153 µmol) in dichloromethane (0.45 mL). Stirring was continued at 18 °C for 10 min then a solution of ptoluenesulfonyl chloride (29 mg, 152 µmol, 1 mol equiv) in dichloromethane (ca. 0.25 mL) was added dropwise over 10 min. After 5 h at 18 °C, the reaction mixture was diluted with dichloromethane (0.4 mL) then filtered through a plug of CeliteTM and the filtrate concentrated under reduced pressure to give a light-yellow oil (67 mg). This was subjected to column chromatography (silica, 1:3:6 v/v/v MeOH/ethyl acetate/hexane elution) and thus affording three fractions, A, B and C.

Concentration of fraction A (R_f =0.4) yielded the C1'S-epimeric form of mono-tosylate 25 (9 mg, 17%) as a clear, colourless oil, $[\alpha]_D$ +26.4 (c 0.64, CHCl₃). Found: M⁺⁺, 350.1182. C₁₈H₂₂O₅S requires M⁺, 350.1188. ¹H NMR (500 MHz, CDCl₃) δ 7.80 (d, J=8.0 Hz, 2H), 7.36 (d, J=8.0 Hz, 2H), 4.17 (dd, J=10.4 and 4.6 Hz, 1H), 4.13 (dd, $J=10.4$ and 2.7 Hz, 1H), 3.56-3.50 (complex m, 1H), 2.77-2.71 (complex m, 1H), 2.49 (dd, $J=5.9$ and 5.3 Hz, 1H), 2.45 (s, 3H), 2.37 (ddd, J=17.8, 2.3 and 1.2 Hz, 1H), 2.26-2.12 (complex m, 2H), 1.94 -1.88 (complex m, 1H), 1.85 (dd, J=9.8 and 5.3 Hz, 1H), 1.81 (d, J=13.4 Hz, 1H), 1.38 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 215.3 (C), 145.1 (C), 132.4 (C), 130.0 (CH), 128.0 (CH), 72.7 (CH2), 71.4 (CH), 56.0 (CH₂), 48.6 (CH₂), 46.3 (C), 43.8 (CH), 42.3 (CH), 38.9 (CH), 34.6 (CH), 25.9 (CH₃), 21.7 (CH₃); IR ν_{max} (KBr) 3418, 2954, 2926, 2870, 1715, 1598, 1453, 1406, 1357, 1309, 1189, 1176, 1098, 1019, 972, 957, 917, 878, 855, 814, 667, 555 cm $^{-1}$; MS (EI, 70 eV) m/z 350 (M $^{+}$; 1%), 165 (31), 155 (33), 136 (100), 105 (45), 93 (51), 91 (80).

Concentration of fraction B (R_f =0.35) afforded a ca. 1:1.5 mixture of the C1'S- and C1'R-epimeric forms of mono-tosylate 25 (29 mg, 54%) as a clear, colourless oil.

Concentration of fraction C (R_f =0.3) gave C1'R-epimeric form of mono-tosylate **25** (3 mg, 6%) as a clear, colourless oil, α _D +25.7 (c 0.35, CHCl3). Found: M^{+•}, 350.1188. C₁₈H₂₂O₅S requires M^{+•}, 350.1188. ¹H NMR (500 MHz, CDCl₃) δ 7.78 (d, J=8.0 Hz, 2H), 7.35 (d, J=8.0 Hz, 2H), 4.06 (dd, J=10.5 and 2.7 Hz, 1H), 3.90 (dd, J=10.5 and 6.3 Hz, 1H), 3.58-3.52 (complex m, 1H), 2.73-2.66 (complex m, 1H), 2.55–2.48 (complex m, 2H), 2.45 (s, 3H), 2.40 (dd, J 17.8 and 2.2 Hz, 1H), 2.18-2.02 (complex m, 3H), 1.99 (dd, $J=9.5$ and 5.2 Hz, 1H), 1.37 (s, 3H), 1.33 (d, J=13.4 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃) d 215.3 (C), 145.1 (C), 132.7 (C), 129.9 (CH), 127.9 (CH), 73.1 (CH2), 71.3 (CH), 56.4 (CH₂), 48.6 (CH₂), 46.6 (C), 43.6 (CH), 42.1 (CH), 39.0 (CH), 35.7 (CH), 25.8 (CH₃), 21.7 (CH₃); IR ν_{max} (KBr) 3420, 2956, 2918, 2870, 2850, 1712, 1598, 1453, 1358, 1312, 1189, 1176, 1097, 972, 957, 946, 918, 880, 853, 815, 667, 555 cm $^{-1}$; MS (EI, 70 eV) m/z 350 (M⁺*, 5%), 165 (47), 136 (100), 93 (68), 91 (95), 43 (67), 32 (48).

3.2.14. Compounds 26 and 27. Method A: A magnetically stirred solution of a ca. 1:1.5 mixture of the C1'S- and C1'R-epimeric forms of mono-tosylate 25 (29 mg, 83 μ mol) in dichloromethane (2.9 mL) was cooled to 0 °C then triethylamine (58 μ L, 414 μ mol, 5 mol equiv) was added followed by DMAP $(834 \text{ µg}, 8.28 \text{ µmol})$ 10 mol %) and benzoyl chloride $(20.7 \mu L, 166 \mu m$ ol, 2 mol equiv). The resulting mixture was allowed to warm to 18 $^\circ\mathsf{C}$ then stirred at this temperature for 14 h before being quenched with sodium hydrogen carbonate (1 mL of a saturated aqueous solution). The separated aqueous phase was extracted with dichloromethane $(5\times2$ mL) then the combined organic phases were dried (magnesium sulfate), filtered and concentrated under reduced pressure. The clear, colourless oil (51 mg) thus obtained was subjected to column chromatography (silica, 1:3:6 v/v/v MeOH/ethyl acetate/ hexane elution) to afford two fractions, A and B.

Concentration of fraction A (R_f =0.5) gave compound 27 (15 mg, 40%) as a clear, colourless oil, [α]_D +14.2 (*c* 0.38, CHCl₃). Found: M⁺⁺, 454.1447. $C_{25}H_{26}O_6S$ requires M⁺, 454.1450. ¹H NMR (500 MHz, CDCl₃) δ 7.92 (dd, J=8.3 and 1.2 Hz, 2H), 7.73 (d, J=8.3 Hz, 2H), 7.59–7.55 (complex m, 1H), 7.42 (dd, J=8.3 Hz, 2H), 7.20 (d, J=8.3 Hz, 2H), 4.98 (m, $J=11.1$ Hz, 1H), 4.44 (dd, $J=11.2$ and 2.6 Hz, 1H), 4.24 (dd, $J=11.2$ and 2.2 Hz, 1H), 3.24 (m, 1H), 2.54 (dd, $J=5.9$ and 5.3 Hz, 1H), 2.40 (dd, J=18.1 and 1.0 Hz, 1H), 2.34 (s, 3H), 2.30 (d, J=18.1 Hz, 1H), 2.20 (ddd, $J=13.2$, 10.7 and 2.0 Hz, 1H), 2.11-2.06 (complex m, 1H), 1.95 (dd, J=9.8 and 5.3 Hz, 1H), 1.55 (d, J=13.2 Hz, 1H), 1.36 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 214.8 (C), 165.5 (C), 144.9 (C), 133.3 (CH), 132.3 (C),129.9 (CH), 129.8 (CH),129.5 (C),128.3 (CH),127.9 (CH), 73.0 (CH) , 69.7 (CH₂), 55.3 (CH₂), 48.7 (CH₂), 46.5 (C), 42.3 (CH), 41.1 (CH), 39.0 (CH), 34.0 (CH), 25.8 (CH₃), 21.6 (CH₃); IR ν_{max} (KBr) 2956, 2927, 1720, 1599, 1452, 1364, 1270, 1190, 1177, 1109, 1097, 1070, 1026, 962, 946, 922, 881, 814, 793, 714, 667, 554 cm $^{-1}$; MS (EI, 70 eV) m/z 454 (Mþ , 4%),149 (18),118 (55),105 (100), 91 (27), 77 (34), 57 (32), 43 (35).

Concentration of fraction B (R_f =0.4) afforded compound 26 (21 mg, 55%) as a white, crystalline solid, mp=134–139 \degree C. Found: M⁺, 454.1448. C₂₅H₂₆O₆S requires M⁺, 454.1450. ¹H NMR (300 MHz, CDCl₃) δ 7.97-7.93 (complex m, 2H), 7.70-7.66 (complex m, 2H), 7.60-7.54 (complex m, 1H), 7.45-7.40 (complex m, 2H), 7.15-7.12 (complex m, 2H), 4.87 (dt, J=11.2 and 2.9 Hz, 1H), 4.31 (dd, J=11.4 and 2.9 Hz, 1H), 4.17 (dd, J=11.4 and 2.9 Hz, 1H), 3.34 (m, 1H), 2.51 (t, $J=5.4$ Hz, 1H), 2.44–2.30 (complex m, partially concealed, 1H), $2.34-2.24$ (complex m, partially concealed, 1H), 2.30 (s, 3H), 2.15-2.07 (complex m, partially concealed, 1H), 2.07 (d, $J=17.0$ Hz, 1H), 1.87 (dd, J=10.2 and 5.4 Hz, 1H), 1.44 (d, J=13.2 Hz, 1H), 1.39 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 213.7 (C), 165.1 (C), 145.0 (C), 133.2 (CH), 132.1 (C), 129.9 (CH), 129.8 (CH), 129.5 (C), 128.2 (CH), 127.8 (CH) , 73.2 (CH), 69.4 (CH₂), 56.2 (CH₂), 47.8 (CH₂), 46.5 (C), 42.1 (CH), 40.9 (CH), 38.9 (CH), 35.6 (CH), 25.8 (CH₃), 21.6 (CH₃); IR ν_{max} (KBr) 2956, 1723, 1599, 1451, 1361, 1312, 1266, 1189, 1176, 1109, 1096, 1069, $1025, 957, 946, 921, 884, 839, 814, 713, 667, 554$ cm⁻¹; MS (EI, 70 eV) m/z 454 (M⁺⁺, 3%), 283 (16), 118 (72), 105 (100), 91 (36), 77 (41).

Method B : A magnetically stirred solution of the $C1/R$ -epimeric form of mono-tosylate 25 (7 mg, 20 μ mol) in dichloromethane (0.7 mL) was treated in the same way as described immediately above. The clear, colourless oil (5 mg) thus obtained was subjected to column chromatography (silica, 1:3:6 v/v/v MeOH/ethyl acetate/hexane elution) and concentration of the appropriate fractions (R_f =0.5) then gave compound 27 (3 mg, 35%) as a white, crystalline solid. This material was identical, in all respects, with an authentic sample.

3.2.15. Compound 28. A solution of diquinane $26(21 \text{ mg}, 46 \text{ µmol})$ in THF/MeOH (0.9 mL of a 2:1 v/v mixture) was cooled to -78 °C and samarium(II) diiodide (0.55 mL of a 0.1 M solution in THF, 55μ mol, 1.2 mol equiv) was added dropwise over 5 min. The resulting mixture was stirred at -78 °C until the initial blue colour associated with the reaction mixture had turned yellow (ca. 25 min) then more samarium(II) diiodide (0.55 mL of a 0.1 M solution in THF, 55 μ mol, 1.2 mol equiv) was added dropwise over 15 min and the reaction mixture warmed to 0° C at which point sufficient additional samarium(II) diiodide was added to re-establish a deep-blue colour. The reaction mixture was then warmed to 18 \degree C and stirred at this temperature until the blue colour had faded. This procedure was repeated twice more at 18 °C until the starting material had been consumed as determined by TLC analysis. The reaction mixture was then quenched with potassium carbonate (1 mL of a saturated aqueous solution) and the separated aqueous phase extracted with diethyl ether $(3\times5 \text{ mL})$ and then with dichloromethane $(3\times5$ mL). The combined organic phases were dried (magnesium sulfate), filtered and concentrated under reduced pressure. The yellow oil thus obtained was subjected to column chromatography (silica, 1:3:6 v/v/v MeOH/ethyl acetate/ hexane elution) and concentration of the appropriate fractions (R_f =0.6) then gave diquinane **28** (11 mg, 52%) as a clear, colourless oil. Found: M⁺, 456.1606. C₂₅H₂₈O₆S requires M⁺, 456.1607. ¹H NMR (300 MHz, CDCl₃) δ 7.94-7.88 (complex m, 2H), 7.74-7.68 (complex m, 2H), $7.62-7.55$ (complex m, 1H), $7.47-7.40$ (complex m, 2H), 7.21-7.15 (complex m, 2H), 5.16-5.06 (m, 1H), 4.22 (dd, $J=11.1$ and 3.4 Hz, 1H), 4.15 (dd, $J=11.1$ and 4.6 Hz, 1H), 2.74-2.57 (complex m, 1H), 2.52 (ddd, J=18.8, 8.8 and 1.2 Hz, 1H), 2.34 (s, 3H), 2.36–2.08 (complex m, 4H), 2.07 (dd, $J=18.8$ and 4.8 Hz, 1H), 1.79 (dd, $J=12.9$ and 8.0 Hz, 1H), 1.41 (dd, $J=12.9$ and 11.1 Hz, 1H), 1.41-1.30 (complex m, 1H), 1.18 (s, 3H); ¹³C NMR (75 MHz, CDCl3) d 219.0 (C), 165.6 (C), 145.0 (C), 133.3 (CH), 132.3 (C), 129.8 (CH), 129.7 (CH), 129.4 (C), 128.4 (CH), 127.8 (CH), 74.3 (CH), 69.5 (CH₂), 52.4 (CH₂), 46.7 (CH), 46.3 (C), 44.2 (CH₂), 42.5 (CH₂), 39.7 (CH), 36.3 (CH₂), 27.7 (CH₃), 21.6 (CH₃); IR ν_{max} (KBr) 3020, 2954, 2918, 2848, 1736, 1727, 1452, 1364, 1269, 1216, 1190, 1177, 1110, 1097, 814, 756, 713, 667, 554 cm $^{-1}$; MS (EI, 70 eV) m/z 456 (M $^{+}$, <1%), 285 (11), 179 (21), 162 (25), 105 (100), 91 (34), 77 (32).

3.2.16. Compound 29. A solution of diquinane $27(15 \text{ mg}, 33 \text{ µmol})$ in THF/MeOH (0.6 mL of a 2:1 v/v mixture) was cooled to -78 °C then samarium(II) diiodide (0.4 mL of a 0.1 M solution in THF, 40 μ mol, 1.2 mol equiv) was added dropwise over 15 min. The resulting mixture was stirred at -78 °C for 20 min then warmed to 0 °C, stirred at this temperature for 3 h then at 18 $^{\circ}$ C for 1 h. After this time, the reaction mixture was re-cooled to -78 °C and additional samarium(II) diiodide $(0.4 \text{ mL of a } 0.1 \text{ M}$ solution in THF, 40μ mol, 1.2 mol equiv) was added dropwise over 2 min. The reaction mixture was stirred at -78 °C for 1 min and then allowed to warm to 18 °C. After 0.5 h at this temperature samarium(II) diiodide (0.4 mL of a 0.1 M solution in THF, 40 μ mol, 1.2 mol equiv) was again added and after 0.5 h at 18 °C the reaction mixture was quenched with potassium carbonate (1 mL of a saturated aqueous solution). The separated aqueous phase was extracted with diethyl ether $(3\times5$ mL) then saturated with sodium chloride and extracted with dichloromethane $(3\times5$ mL). The combined organic phases were dried (magnesium sulfate), filtered and concentrated under reduced pressure. The resulting yellow oil was subjected to column chromatography (silica, 1:3:6 v/v/v MeOH/ethyl acetate/hexane elution) and concentration of the appropriate fractions (R_f =0.5) then gave diquinane **29** (3 mg, 21%) as a clear, colourless oil. Found: $(M+Na)^+$, 479.1500. C₂₅H₂₈O₆S requires $(M+Na)^+$, 479.1504. ¹H NMR (500 MHz, CDCl₃): δ 7.93–7.89 (complex m, 2H), $7.73-7.69$ (complex m, 2H), $7.61-7.56$ (complex m, 1H), $7.46-7.42$ (complex m, 2H), 7.20-7.16 (complex m, 2H), 5.12 (ddd, J =7.8, 4.5 and 3.4 Hz, 1H), 4.21 (dd, J=11.1 and 3.4 Hz, 1H), 4.17 (dd, J=11.1 and 4.5 Hz, 1H), 2.62–2.72 (complex m, 1H), 2.53 (ddd, J=19.0, 9.3 and 1.5 Hz, 1H), 2.34 (s, 3H), 2.34–2.14 (complex m, 4H), 2.05 (ddd, J=19.0, 4.6 and 1.5 Hz, 1H), 1.84 (dd, J=13.2 and 7.8 Hz, 1H), 1.58 (dd, J=13.2 and 10.7 Hz, 1H), 1.22-1.14 (complex m, 1H), 1.18 (s, 3H); ¹³C NMR $(125 MHz, CDCl₃)$ δ 219.0 (C), 165.6 (C), 145.0 (C), 133.4 (CH), 132.4 (C), 129.8 (CH),129.7 (CH),129.4 (C),128.4 (CH),127.9 (CH), 74.2 (CH), 69.5 $(CH₂), 52.4 (CH₂), 46.7 (CH), 46.2 (C), 44.4 (CH₂), 42.4 (CH₂), 39.7 (CH),$ 36.3 (CH₂), 27.7 (CH₃), 21.6 (CH₃); IR ν_{max} (KBr) 2953, 1738, 1722, 1451, 1364, 1269, 1190, 1177, 1109, 1097, 1070, 1026, 976, 942, 815, 714, 667, 554 cm⁻¹; MS (ESI, +ve) m/z 495 [(M+K)⁺, 7%], 479 [(M+Na)⁺, 100], 455 (38), 285 (93), 206 (35), 163 (50), 135 (35), 105 (69).

3.2.17. Compound 32. A magnetically stirred solution of diquinane **28** (10 mg, 22 μ mol) in THF (0.2 mL) was cooled to -78 °C then LiHMDS (26 μ L of a 1 M solution in THF, 26 μ mol, 1.2 mol equiv) added dropwise over 5 min. After 0.5 h the reaction mixture was warmed to 0 \degree C and then, after 1 h, to 18 \degree C. After 5 h at this temperature the reaction mixture was quenched with water (0.2 mL) and the separated aqueous phase washed with dichloromethane $(5\times1$ mL). The combined organic layers were then dried (magnesium sulfate), filtered and concentrated under reduced pressure. The resulting light-yellow oil was subjected to column chromatography (silica, 1:3:6 v/v/v MeOH/ethyl acetate/hexane elution) and thereby affording two fractions, A and B.

Concentration of fraction A (R_f =0.7) gave compound 32 (3 mg, 53% at 91% conversion) as a white, crystalline solid, mp= $128-133$ °C. Found: M^{+•}, 284.1412. C₁₈H₂₀O₃ requires M^{+•}, 284.1412. ¹H NMR (300 MHz, CDCl₃) δ 8.04–7.99 (complex m, 2H), 7.58–7.52 (complex m, 1H), 7.47-7.40 (complex m, 2H), 4.73 (t, J=8.1 Hz, 1H), 2.56 (dd, $J=18.3$ and 7.8 Hz, 1H), 2.46–2.17 (complex m, 4H), 2.10 (d, $J=9.1$ Hz, 1H), 1.96 (d, J=12.3 Hz, 1H), 1.76 (dt, J=13.7 and 9.1 Hz, 2H), 1.45 (dd, $J=12.3$, 4.5 and 1.1 Hz, 1H), 1.26 (s, 3H), 1.19–1.11 (complex m, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 222.6 (C), 165.8 (C), 132.9 (CH), 130.5 (C), 129.5 (CH), 128.3 (CH), 75.8 (CH), 50.9 (CH), 47.0 (C), 44.3 (CH2), 43.1 (CH), 41.3 (CH), 38.4 (CH₂), 36.0 (CH₂), 25.8 (CH₂), 25.0 (CH₃); IR ν_{max} (KBr) 2930, 1732, 1717, 1451, 1336, 1313, 1275, 1258, 1179, 1143, 1112, 1068, 1024, 1003, 981, 710 cm⁻¹; MS (EI, 70 eV) m/z 284 [M⁺⁺, 3%], 163 (45), 162 (89), 134 (52), 106 (44), 105 (100), 93 (89), 92 (63), 77 (83). A sample of this material was recrystallised (ethyl acetate/ hexane) to provide a crystal suitable for X-ray analysis. Details of this analysis are presented below and in [Figure 2](#page-4-0).

Concentration of fraction B (R_f =0.6) gave starting material 28 (1 mg, 9% recovery) as a clear, colourless oil that was identical, in all respects, with an authentic sample.

3.3. Single-crystal X-ray analysis of compounds 12 and 32

3.3.1. Data for compound 12. $C_{14}H_{20}O_5$, M=268.31, T=200 K, monoclinic, space group $P2_1$, Z=4, a=14.7414(4), b=6.6985(2), $c=15.2998(5)$ Å, $\beta=111.7498(19)$; $V=1403.23(8)$ Å³, $D_x=1.270$ $g \text{ cm}^{-3}$, 2697 unique data (2 θ_{max} =50°), R=0.0307 [for 2192 reflections with $I > 1.5\sigma(I)$; Rw=0.0376 (all data), S=1.1764.

3.3.2. Data for compound 32. $C_{18}H_{20}O_3$, M=284.36, T=200 K, monoclinic, space group $P2_1$, $Z=2$, $a=6.8933(3)$, $b=7.4523(2)$, c=14.8977(6) \AA , β =100.166(2); V=753.29(5) \AA ³, D_x=1.254 g cm⁻³, 1439 unique data ($2\theta_{\rm max}$ =50°), R=0.035 [for 1084 reflections with $I > 2.0\sigma(I)$; Rw=0.081 (all data), S=0.82.

3.3.3. Structure determination. Images were measured on a Nonius Kappa CCD diffractometer (Mo Ka, graphite monochromator, λ =0.71073 Å) and data extracted using the DENZO package.³⁵

Structure solution was by direct methods (SIR92).³⁶ The structures of compounds 12 and 32 were refined using the CRYSTALS program package.37 Crystallographic data (excluding structure factors) have been deposited at the Cambridge Crystallographic Data Centre (CCDC nos. 763134 and 760492 for compounds 12 and 32, respectively). These data can be obtained free-of-charge via www.ccdc. cam.ac.uk/data_request/cif, by emailing data_request@ccdc.cam.ac. uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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References and notes

- 1. Fusetani, N.; Wolstenholme, H. J.; Matsunaga, S.; Hirota, H. Tetrahedron Lett. 1991, 32, 7291.
- 2. (a) Burreson, B. J.; Scheuer, P. J.; Finer, J.; Clardy, J. J. Am. Chem. Soc. 1975, 97, 4763; (b) Hagadone, M. R.; Burreson, B. J.; Scheuer, P. J.; Finer, J. S.; Clardy, J. Helv. Chim. Acta 1979, 62, 2484; (c) Fusetani, N.; Wolstenholme, H. J.; Matsunaga, S. Tetrahedron Lett. 1990, 31, 5623.
- 3. Karuso, P.; Poiner, A.; Scheuer, P. J. J. Org. Chem. 1989, 54, 2095.
- 4. (a) Hagadone, M. R.; Scheuer, P. J.; Holme, A. J. Am. Chem. Soc. 1984, 106, 2447; (b) Karuso, P.; Scheuer, P. J. J. Org. Chem. 1989, 54, 2092.
- 5. (a) Dumdei, E. J.; Flowers, A. E.; Garson, M. J.; Moore, C. J. Comp. Biochem. Physiol. 1997, 118A, 1385; (b) Simpson, J. S.; Garson, M. J. Tetrahedron Lett. 2001, 42, 4267; (c) Simpson, J. S.; Garson, M. J. Org. Biomol. Chem. 2004, 2, 939; (d) Garson, M. J.; Simpson, J. S. Nat. Prod. Rep. 2004, 21, 164 and references cited therein.
- 6. The absolute stereochemistry of 2-isocyanoallopupukeanane (1) has not been determined although it is presumed to be as illustrated because of its likely biogenetic relationship to 9-isocyanopupukeanane (3) for which the absolute configuration has been established (see Ref. 2b).
- 7. Total and formal total syntheses of (\pm) -2: (a) Corey, E. J.; Ishiguro, M. Tetrahedron Lett. 1979, 20, 2745; (b) Fráter, G.; Wenger, J. Helv. Chim. Acta 1984, 67, 1702; (c) Jaafar, A.; Alilou, E. H.; Réglier, M.; Waegell, B. Tetrahedron Lett. 1991, 32, 5531; (d) Chang, N.-C.; Chang, C.-K. J. Org. Chem. 1996, 61, 4967; (e) Srikrishna, A.; Vijaykumar, D.; Sharma, G. V. R. Tetrahedron Lett. 1997, 38, 2003; (f) Kaliappan, K.; Subba Rao, G. S. R. J. Chem. Soc., Perkin Trans. 1 1997, 3393; (g) Gutke, H.-J.; Oesterreich, K.; Spitzner, D.; Braun, N. A. Tetrahedron 2001, 57, 997; formal total synthesis of $(-)$ -2: (h) Brown, M. K.; Corey, E. J. Org. Lett. 2010, 12, 172; total and formal total syntheses of (\pm) -3: (i) Corey, E. J.; Behforouz, M.; Ishiguro, M. J. Am. Chem. Soc. **1979**, 101, 1608; (j) Yamamoto, H.; Sham, H. L. J. Am.
Chem. Soc. **1979**, 101, 1609; (k) Schiehser, G. A.; White, J. D. J. Org. Chem. **1980**, 45, 1864; (l) Piers, E.; Winter, M. Liebigs Ann. Chem. 1982, 973; (m) Hsieh, S.-L.; Chiu, C.-T.; Chang, N.-C. J. Org. Chem. 1989, 54, 3820; formal total synthesis of $(+)$ -3: (n) Srikrishna, A.; Reddy, T. J. J. Chem. Soc., Perkin Trans. 1 1997, 3293; a total synthesis of (\pm) -4 has also been reported: (o) Ho, T.-L.; Jana, G. H. J. Org. Chem. 1999, 64, 8965.
- 8. (a) Ho, T.-L.; Kung, L.-R. Org. Lett. 1999, 1, 1051; (b) Ho, T.-L.; Kung, L.-R.; Chein, R.-J. J. Org. Chem. 2000, 65, 5774.
- 9. Srikrishna, A.; Satyanarayana, G. Tetrahedron Lett. 2006, 47, 367.
- 10. (a) Banwell, M. G.; Edwards, A. J.; Harfoot, G. J.; Jolliffe, K. A. Tetrahedron 2004, 60, 535; (b) Banwell, M. G.; Austin, K. A. B.; Willis, A. C. Tetrahedron 2007, 63, 6388; (c) Reekie, T. A.; Austin, K. A. B.; Banwell, M. G.; Willis, A. C. Aust. J. Chem. 2008, 61, 94.
- 11. Singh, V. In CRC Handbook of Organic Photochemistry and Photobiology, 2nd ed.; Horspool, W. M., Lenci, F., Eds.; CRC LLC: Boca Raton, FL, 2004; pp 78/1-78/34.
- 12. See, for example: (a) Banwell, M. G.; Darmos, P.; Hockless, D. C. R. Aust. J. Chem. 2004, 57, 41; (b) Banwell, M. G.; McLeod, M. D.; Riches, A. G. Aust. J. Chem. 2004, 57, 53.
- 13. Compound 8 can be obtained from Questor, Queen's University of Belfast, Northern Ireland. Questor Centre Contact Page: http://questor.qub.ac.uk/ newsite/contact.htm (accessed 11.01.10). For reviews on methods for generating cis-1,2-dihydrocatechols by microbial dihydroxylation of the corresponding aromatics, as well as the synthetic applications of these metabolites, see: (a) Hudlicky, T.; Gonzalez, D.; Gibson, D. T. Aldrichimica Acta 1999, 32, 35; (b) Banwell, M. G.; Edwards, A. J.; Harfoot, G. J.; Jolliffe, K. A.; McLeod, M. D.; McRae, K. J.: Stewart, S. G.; Vögtle, M. Pure Appl. Chem. 2003, 75, 223; (c) Johnson, R. A. Org. React. 2004, 63, 117; (d) Hudlicky, T.; Reed, J. W. Synlett 2009, 685.
- 14. Hudlicky, T.; Luna, H.; Barbieri, G.; Kwart, L. D. J. Am. Chem. Soc. 1988, 110, 4735.
- 15. Evans, D. A.; Scott, W. L.; Truesdale, L. K. Tetrahedron Lett. 1972, 13, 121.
- 16. (a) Crabtree, S. R.; Mander, L. N.; Sethi, S. P. Org. Synth. 1992, 70, 256; (b) Bissember, A. C. Synlett 2009, 681.
- 17. The initial assignment of stereochemistry in these systems was based upon the magnitude of the vicinal couplings between H8 and H9 and the observation (or otherwise) of NOEs between H8 and/or H9 and those resonances due to the oxymethine protons associated with the acetonide residue.
- 18. Crossland, R. K.; Servis, K. L. J. Org. Chem. 1970, 35, 3195.
- 19. Parikh, J. R.; Doering, W. v. E J. Am. Chem. Soc. 1967, 89, 5505.
- 20. This protocol is based on one first described by Ma and Bobbitt (Ma, Z.; Bobbitt, J. M. J. Org. Chem. 1991, 56, 6110). For an example of a related oxidation see Ref. 10b.
- 21. Molander, G. A.; Hahn, G. J. Org. Chem. 1986, 51, 1135.
- 22. VanRheenen, V.; Kelly, R. C.; Cha, D. Y. Tetrahedron Lett. 1976, 17, 1973.
- 23. Kolb, H. C.; VanNieuwenhze, M. S.; Sharpless, K. B. Chem. Rev. 1994, 94, 2483.
- 24. Guillaume, M.; Lang, Y. Tetrahedron Lett. 2010, 51, 579 and references cited therein.
- 25. For examples of the reductive-ring opening of cyclopropyl ketones using samarium(II) diiodide see: Batey, R. A.; Motherwell, W. B. Tetrahedron Lett. 1991, 32, 6649.
- 26. For examples of the exploitation of samarium enolates in synthesis see: (a) Nakamura, Y.; Takeuchi, S.; Ohgo, Y.; Yamaoka, M.; Yoshida, A.; Mikami, K. Tetrahedron 1999, 55, 4595; (b) Concellón, J. M.; Rodríguez-Solla, H.; Simal, C. Adv. Synth. Catal. 2009, 351, 1238.
- 27. See, for example: Carey, F. A.; Sundberg, R. J. Advanced Organic Chemistry: Part B, 4th ed.; Springer Science: New York, NY, 2001, pp 5-10.
- 28. Allen, C. C. R.; Boyd, D. R.; Dalton, H.; Sharma, N. D.; Brannigan, I.; Kerley, N. A.; Sheldrake, G. N.; Taylor, S. C. J. Chem. Soc., Chem. Commun. 1995, 117.
- 29. Austin, K. A. B.; Elsworth, J. D.; Banwell, M. G.; Willis, A. C. Org. Biomol. Chem. 2010, 8, 751.
- 30. (a) Cox, P. J.; Simpkins, N. S. Tetrahedron: Asymmetry 1991, 2, 1; (b) Koga, K. Pure Appl. Chem. 1994, 66, 1487; (c) O'Brien, P. J. Chem. Soc., Perkin Trans. 1 1998, 1439; (d) Plaquevent, J.-C; Perrard, T.; Cahard, D. Chem.—Eur. J 2002, 8, 3300.
- 31. For the application of such a desymmetrisation process within a diquinane system see: (a) Izawa, H.; Shirai, R.; Kawasaki, H.; Kim, H.; Koga, K. Tetrahedron Lett. 1989, 30, 7221; (b) Leonard, J.; Ouali, D.; Rahman, S. K. Tetrahedron Lett. 1990, 31, 739; (c) Leonard, J.; Hewitt, J. D.; Ouali, D.; Rahman, S. K.; Simpson, S. J.; Newton, R. F. Tetrahedron: Asymmetry 1990, 1, 699.
- 32. Still, W. C.; Kahn, M.; Mitra, A. J. Org. Chem. 1978, 43, 2923.
- Pangaborn, A. B.; Giardello, M. A.; Grubbs, R. H.; Rosen, R. K.; Timmers, F. J. Organometallics 1996, 15, 1518.
- 34. This ca. 4:1 mixture of alcohols 12 and C8,C9-di-epi-12 was generated by combining material obtained from the reduction of ketone 11 and the epimerisation of compound C9-epi-12.
- 35. DENZO-SMN Otwinowski, Z.; Minor, W. Processing of X-ray diffraction data collected in oscillation mode. In Macromolecular Crystallography, Part A; Carter, C. W., Jr., Sweet, R. M., Eds.; Methods in Enzymology; Academic: New York, NY, 1997; Vol. 276, pp 307-326.
- 36. SIR92 Altomare, A.; Cascarano, G.; Giacovazzo, C.; Guagliardi, A.; Burla, M. C.; Polidori, G.; Camalli, M. J. Appl. Crystallogr. 1994, 27, 435.
- 37. Betteridge, P. W.; Carruthers, J. R.; Cooper, R. I.; Prout, K.; Watkin, D. J. J. Appl. Crystallogr. 2003, 36, 1487.